

Tree responses to rising CO₂ in field experiments: implications for the future forest

R. J. NORBY,¹ S. D. WULLSCHLEGER,¹ C. A. GUNDERSON,¹ D. W. JOHNSON,² & R. CEULEMANS³

¹Environmental Sciences Division, Oak Ridge National Laboratory, Building 1059, PO Box 2008, Oak Ridge, TN 37831-6422, USA,

²Desert Research Institute, Reno, Nevada, USA and ³Department of Biology, University of Antwerpen (UIA), B-2610 Wilrijk, Belgium

ABSTRACT

The need to assess the role of forests in the global cycling of carbon and how that role will change as the atmospheric concentration of CO₂ increases has spawned many experiments over a range of scales. Experiments using open-top chambers have been established at many sites to test whether the short-term responses of tree seedlings described in controlled environments would be sustained over several growing seasons under field conditions. Here we review the results of those experiments, using the framework of the interacting cycles of carbon, water and nutrients, because that is the framework of the ecosystem models that are being used to address the decades-long response of forests.

Our analysis suggests that most of what was learned in seedling studies was qualitatively correct. The evidence from field-grown trees suggests a continued and consistent stimulation of photosynthesis of about 60% for a 300 p.p.m. increase in [CO₂], and there is little evidence of the long-term loss of sensitivity to CO₂ that was suggested by earlier experiments with tree seedlings in pots. Despite the importance of respiration to a tree's carbon budget, no strong scientific consensus has yet emerged concerning the potential direct or acclimation response of woody plant respiration to CO₂ enrichment. The relative effect of CO₂ on above-ground dry mass was highly variable and greater than that indicated by most syntheses of seedling studies. Effects of CO₂ concentration on static measures of response are confounded with the acceleration of ontogeny observed in elevated CO₂. The trees in these open-top chamber experiments were in an exponential growth phase, and the large growth responses to elevated CO₂ resulted from the compound interest associated with an increasing leaf area. This effect cannot be expected to persist in a closed-canopy forest where growth potential is constrained by a steady-state leaf area index. A more robust and informative measure of tree growth in these experiments is the annual increment in wood mass per unit leaf area, which increased 27% in elevated CO₂. There is no support for the conclusion from many studies of seedlings that root-to-shoot ratio is increased by elevated CO₂; the production of fine roots may be enhanced, but it is not clear

that this response would persist in a forest. Foliar nitrogen concentrations were lower in CO₂-enriched trees, but to a lesser extent than was indicated in seedling studies and only when expressed on a leaf mass basis. The prediction that leaf litter C/N ratio would increase was not supported in field experiments. Also contrasting with seedling studies, there is little evidence from the field studies that stomatal conductance is consistently affected by CO₂; however, this is a topic that demands more study.

Experiments with trees in open-top chambers under field conditions have provided data on longer-term, larger-scale responses of trees to elevated CO₂ under field conditions, confirmed some of the conclusions from previous seedling studies, and challenged other conclusions. There remain important obstacles to using these experimental results to predict forest responses to rising CO₂, but the studies are valuable nonetheless for guiding ecosystem model development and revealing the critical questions that must be addressed in new, larger-scale CO₂ experiments.

Key-words: atmospheric carbon dioxide; forests; global change; open-top chambers; trees

TREES, FORESTS, AND CO₂: A PROBLEM OF SCALE

Presenting the experimental evidence on the response of trees to elevated CO₂ is primarily a problem of scale. The rationale for most of the experiments that have been conducted under the global change umbrella is the need to assess the role of forests in the global cycling of carbon and how that role will change as the atmosphere becomes progressively enriched with CO₂. But the scale of most experiments is not that of the forest. Even the longest-duration CO₂ experiments represent only a small fraction of the life of a tree. No matter how well an experiment with a tree seedling is conducted and how well the data are summarized, the effort is of little use if there is no framework for interpreting the results in the context of the decades-long responses of forest trees and the forest ecosystem to rising CO₂. Our challenge is to find an appropriate framework.

Experimental research on tree responses to CO₂ over the past two decades is characterized by a gradual increase in the scale and complexity of investigations. Following a time-honoured paradigm of scientific inquiry, simple

Correspondence: Richard J. Norby. Fax: 1 423 576 9939; e-mail: rjn@ornl.gov

experiments gave rise to new insights, new questions and new hypotheses to test. For example, experiments with tree seedlings demonstrated that growth can increase in elevated CO₂ even under nutrient-limited conditions and that photosynthesis usually increases, but that photosynthetic capacity may decline, foliar N values are reduced, and stomatal closure reduces water use. But do foliar N and photosynthetic capacity decline when the roots are not constrained by pots? Does growth continue to be stimulated by elevated CO₂ over several growing seasons under field conditions of multiple, fluctuating environmental variables? These critical questions, and many others, could be answered only in experiments in which the trees were rooted in unconstrained soil and grown in chambers large enough to accommodate several years' worth of growth. Hence, experiments using open-top chambers, an exposure technology proven in air-pollution research and adapted for elevated CO₂ studies (Rogers, Heck & Heagle 1983), were established around the world. These experiments used different species under different conditions to address different questions. Together, they provide a wealth of data and understanding about how forest trees will respond to the inexorably increasing CO₂ concentration in the atmosphere. But we will inevitably find that the data are inadequate, the experimental approaches flawed, and the prospects for understanding forest growth and metabolism in the future still unfulfilled. We should, however, take some lessons from these experiments, and form new questions, new hypotheses, to guide the next wave of larger-scale, longer-term experiments geared toward understanding forest response to global change. Larger-scale experiments, which will inevitably be more expensive and more difficult, will be most profitable if guided by testable hypotheses based on our best and most current understanding. That is the spirit in which we present this review of tree responses to elevated CO₂.

We start with a well established body of published studies, describing many of the mechanisms by which tree species respond to elevated CO₂ and the expression of those mechanisms in the growth of young seedlings. Even the reviews on tree responses are too numerous to list, but much of the progress in the field can be tracked through Oechel & Strain (1985), Eamus & Jarvis (1989), and Ceulemans & Mousseau (1994). Other reviews have focussed on specific processes such as photosynthesis (Gunderson & Wullschleger 1994), below-ground processes (Norby 1994) and nitrogen concentrations (McGuire, Melillo & Joyce 1995; Cotrufo, Ineson & Scott 1998), or have compiled the many data sets in a format useful for modellers (Wullschleger, Post & King 1995a; Curtis 1996; Wullschleger, Norby & Gunderson 1997a; Curtis & Wang 1998). Collectively, these reviews indicate that with an increase in CO₂ concentration to \approx 650–700 p.p.m., photosynthesis and dry mass increase, and foliar nutrient concentrations decline. Meta-analysis (Curtis & Wang 1998) has indicated that field-grown trees may respond differently from trees in pots, although those conclusions were necessarily tentative because of sparse

data sets. Wullschleger *et al.* (1997a) concluded that data derived from short-term experiments may at best set upper bounds on how the larger biosphere might respond to long-term increases in CO₂. As long as our ultimate interest is the long-term response of trees in a forest, then it is critical that we ensure that our projections into the future are based on the most relevant data available. The number of new field studies has increased rapidly, and it is now possible to look at them separately from the large pool of previous CO₂ studies.

Ecosystem models provide a useful organizing tool for summarizing tree responses to global change. The decades-long response of forests can be addressed only through models, and the response of the tree is necessarily a dominant factor in such models. For the models to have a flexible, predictive value, they must contain explicit descriptions of the processes on which the global change factor acts (Jarvis 1995). The mathematical expressions of CO₂ responses in these models are developed on the basis of biological principles, intuition or a qualitative assessment of experimental results. The opportunity to shape and constrain modelling efforts with experimental data is attractive, but in doing so we must be careful that the data we use really are appropriate. Most ecosystem models are organized around the intersecting cycles of carbon, water and nutrients—collectively, biogeochemical cycling. The effects of CO₂ are included primarily in five ways: effects on stomatal conductance and water-use efficiency; photosynthesis and respiration; carbon allocation and growth; plant structure and phenology; and plant nutrient concentrations (Mooney *et al.* 1999). Because of the high priority assigned to the provision of data on tree responses to elevated CO₂ that can guide models, our discussion will be organized around the cycles of carbon, nutrients, and water.

THE DATABASE OF FIELD-GROWN TREES IN ELEVATED CO₂

The primary database we draw on for this review and synthesis is summarized in Table 1. These are the experiments with tree species planted in the ground and exposed with replication to elevated CO₂ for at least one growing season. Additional field experiments have been conducted with mature trees for shorter durations (Wong & Dunin 1987), without replication (Surano *et al.* 1986), with branch bags to enrich only isolated branches (Teskey 1995), with potted seedlings (Murray *et al.* 1994; El Kohen, Venet & Mousseau 1993), or with constructed microcosms (Körner & Arnone 1992; Overdieck 1993; Hättenschwiler & Körner 1998). Such experiments certainly can be valuable and answer specific questions, and our strict criteria are not meant to denigrate other approaches. There have been many reviews and syntheses of tree responses to elevated CO₂ that encompassed the entire data set. Some of the conclusions from these reviews may be challenged because of the many confounding factors related to experimental approach. This synthesis will be based on a much more limited data set, but perhaps a data set that has fewer confounding factors.

The experiments listed in Table 1 were designed with different objectives and different limitations. Simply stated, however, an important objective of all such experiments has been to determine if the responses to elevated CO₂ measured in young seedlings in greenhouses and growth chambers are sustained over several growing seasons under field conditions (Norby, Wullschleger & Gunderson 1996). In all cases compromises were necessarily made in the size and kind of exposure system, the number of replicates, the nature of the initial plant material, and management of the soil environment. Most of the experiments were conducted in open-top chambers, which permitted the plants to be planted directly in the soil and grown under conditions near-ambient except for the introduction of additional CO₂ into the atmosphere. However, open-top chambers attenuate light and elevate temperature, unless they are specifically engineered to control temperature (Norby *et al.* 1997) or are used in an understory (Cipollini, Drake & Whigham 1993). Hence, they cannot be considered to provide true ambient conditions. Furthermore, most of these experiments were not conducted within a true forest setting, and the soil and light conditions (particularly side light) were not typical of the forest (Körner 1995). The duration of most experiments was limited by the size of the chambers. Most of the species that have been investigated are from the North Temperate or Mediterranean forests and encompass a broad range of deciduous, broadleaf evergreen, and coniferous species. Table 1 is not necessarily complete, and experiments under way or recently completed will augment this data base with different species and different interacting variables.

It is especially important to recognize that in none of these experiments was the experimental unit a forest ecosystem. In most cases, the experimental unit was an isolated tree or group of young trees. The objectives of the experiments cannot be to measure the response of forest ecosystems to elevated CO₂, but instead to measure some of the important component processes with the intention that those measurements will provide some insight to the higher-scale processes of interest. This is the perspective we must maintain as we interpret the experimental results.

CARBON CYCLING

The central focus of most of the studies in Table 1, and research on elevated CO₂ effects in general, is the carbon cycle. Will increased tree growth in elevated CO₂ cause a higher fraction of fossil-fuel-derived carbon to be stored in the biosphere, thereby slowing the increase in the atmospheric concentration and forestalling climatic change? Will increased carbon assimilation by trees enhance the flux of carbon to long-lived soil carbon pools? Carbon cycle studies begin with the biochemical processes of photosynthesis and plant respiration and increase in scale to that of whole-plant growth and allocation. At higher scales there are important interactions with nutrient and water cycles, which are critical to whole-ecosystem assessments.

Photosynthesis

The first physiologically meaningful contact between plant and atmosphere takes place at the leaf, and most subsequent effects of increasing CO₂ concentration are linked to changes in CO₂ assimilation. Because of this, a great deal of attention has focused on leaf-level photosynthetic responses to CO₂ enrichment. The undisputed response to increasing [CO₂] is an increase in photosynthesis, but a host of questions have arisen concerning longer-term effects after growth and development at a higher CO₂ concentration. The key question, relative to global change impacts on forests, is how much photosynthesis will increase as atmospheric CO₂ concentrations rise, and what bearing this will have on higher-level processes. The answer may (or may not) be complicated by interactions with other environmental gradients, and may vary within the canopy, seasonally or between species. In addition, because some very early CO₂ enrichment experiments reported complete losses of photosynthetic enhancement after extremely short exposure times (e.g. days to weeks; reviewed for crops by Cure & Acock 1986), there has been a particular focus on detecting and explaining possible decreases in photosynthetic stimulation over time.

The first part of the question, the magnitude of photosynthetic response to CO₂ that can be sustained over a season or several seasons, can be addressed by comparing assimilation at the growth CO₂ concentration, typically measured on single leaves at light saturation. In trees growing outdoors, rooted in the ground, these rates were almost always higher in elevated CO₂, regardless of the duration of the study. Photosynthesis was stimulated 40–80% in most of the studies reviewed here, although in several cases the enhancement was substantially greater (Table 2, Fig. 1a). The mean enhancement of 66% (geometric mean 63%) is greater, and the variability is less, than that reported in a previous review of tree responses (44%, Gunderson & Wullschleger 1994), at which time most available data were from experiments with potted material, and encompassed a wider range of [CO₂].

The field experiments have been useful for describing how other environmental variables could modify the photosynthetic responses to CO₂. The photosynthetic response might be reduced by nutrient deficiency (Eamus & Jarvis 1989; Tissue, Thomas & Strain 1993; Curtis *et al.* 1994; Sage 1994), or conversely, enhanced in combination with other stresses (Long 1991; Idso & Idso 1994), or unaffected by stress (Curtis & Wang 1998). Conflicting interactions between CO₂ and nitrogen concentration have also been related to secondary effects of nutrient supply on growth and sink strength (Pettersson & McDonald 1994), which could complicate the interpretation of experimental results. In three field experiments in which nutrients were deliberately manipulated (Table 2), season-long enhancements were greater in the high-nutrient treatments (Curtis *et al.* 1995; Kubiske *et al.* 1997) or increased after nutrients were added (Curtis *et al.* 1994), but photosynthesis was enhanced by 40–62% even in the lower-nutrient treatments,

Table 1. Protocol of replicated experiments in which whole trees were exposed to elevated CO₂ under field conditions

	CO ₂ levels*	Interacting factors	Reps	Plants chamber ⁻¹	Area (m ²)	Duration (seasons)	Initial plant material	Cultural treatments	Reference
Angiosperms									
<i>Acer rubrum</i>	2	Temperature	3	20	7.1	4	1-year-old seedlings from nursery	None	Norby <i>et al.</i> 1997, Norby <i>et al.</i> 1998
<i>Acer saccharum</i>									
<i>Acer rubrum</i>	2	Shade	6	6	0.5	1	1-year-old seedlings from nursery	Soil boxes	Kubiske & Pregitzer 1996
<i>Betula papyrifera</i>									
<i>Quercus rubra</i>									
<i>Acer saccharum</i>	2	Defoliation	4	18	10.2	2	4-year-old saplings from nursery; seedlings from seed	Irrigated	Roth <i>et al.</i> 1998
<i>Populus tremuloides</i>									
<i>Alnus glutinosa</i>	2		5	5	0.5	1	3-month-old seedlings	Mixed soil in open-bottom boxes	Vogel & Curtis 1995
<i>Betula pendula</i>	2		6	1	1.8	4	Seedlings grown from seed	Irrigated, fertilized	Rey & Jarvis 1997
<i>Citrus aurantium</i>	2		2	2	13.8–31.6	8 [†]	Seedlings	Irrigated, fertilized	Idso & Kimball 1997
<i>Lindera benzoin</i>	2		3	30	12	2	Naturally occurring ramets in forest understory	None	Cipollini <i>et al.</i> 1993
<i>Liriodendron tulipifera</i>	3		2	5	7.1	2.5	Seedlings grown from seed	None	Norby <i>et al.</i> 1992
<i>Mangifera indica</i>	2		2	24		3 wet 2 dry	Grafted seedlings	Imported top soil; irrigated, fertilized	Goodfellow <i>et al.</i> 1997
<i>Populus clones</i>	2	Clones of different growth rate	2	15	7	2	Unrooted hardwood cuttings	Fertile horticultural soil; irrigated, fertilized	Ceulemans <i>et al.</i> 1996
<i>Populus deltoides</i> × <i>P. nigra</i>	2	Soil fertility	5	5	0.5	1	Cuttings	Planted in open-bottom root boxes; irrigated	Curtis <i>et al.</i> 1995
<i>Populus grandidentata</i>	2		4	9	0.25	1	Rooted cuttings	Homogenized subsoil in open-bottom boxes with added N; irrigated	Zak <i>et al.</i> 1993
<i>Populus tremuloides</i>	2	Clones with different O ₃ sensitivity	3	12	?	3	3-month-old cuttings	Twice-ambient ozone; irrigated	Karnosky <i>et al.</i> 1998
<i>Quercus alba</i>	3		2	5	7.1	4	Seedlings grown from seed	None	Norby <i>et al.</i> 1996
<i>Quercus ilex</i>	2		3	2	12.6	3	Natural community	None	Scarascia-Mugnozza <i>et al.</i> 1996
<i>Quercus sp.</i>									
<i>Serenoa repens</i>	2		3		4.3	1	Natural community, resprouting after cut	None	Day <i>et al.</i> 1996

Table 1. Continued.

	CO ₂ levels*	Interacting factors	Reps	Plants chamber ⁻¹	Area (m ²)	Duration (seasons)	Initial plant material	Cultural treatments	Reference
<i>Gymnosperms</i>									
<i>Picea abies</i>	2		4	1		4	10-year-old trees in situ	None	Dvorak & Oplustilova 1997
<i>Picea sitchensis</i>	2	Nitrogen				2	3.5-year-old seedlings		Lee <i>et al.</i> 1998;
<i>Pinus eldarica</i>	4		2	2	9	2	40-cm tall seedlings from nursery	Irrigated and fertilized	Murray & Ceulemans 1998 Idso & Kimball 1994
<i>Pinus ponderosa</i>	3	Nitrogen	3	21	11.2	6	Seed	Irrigated	Johnson <i>et al.</i> 1996
<i>Pinus radiata</i>	2		4	8	17.3	1	Plantlets from tissue culture	Irrigated and fertilized	Thomas <i>et al.</i> 1996
<i>Pinus sylvestris</i>	2	Temperature	4	1	6.2	4	20–25-year-old trees in situ	Irrigated	Wang <i>et al.</i> 1995
<i>Pinus sylvestris</i>	2	Ozone	4	1	3.2	3	30-year-old trees in situ	Imported forest soil;	Kellomäki & Wang 1997
<i>Pinus sylvestris</i>	2		2	11	7		3-year-old seedlings	no soil amendments	Janssens <i>et al.</i> 1998
<i>Pinus taeda</i>	2		3	24	7.1–19.6	4	Seed	Mixed soil	Tissue <i>et al.</i> 1997
<i>Pseudotsuga menziesii</i>	2	Temperature	3	14	2	3.5	2-year-old seedlings	Imported forest soil in lysimeters; irrigated	Guak <i>et al.</i> 1998
Mixtures									
<i>Fagus sylvatica</i>									
<i>Picea abies</i>	2	Wet N deposition; soil type	4	32	6.7	1 [†]	2–3-year old seedlings and clonal cuttings	Imported forest soil	Egli & Körner 1997
<i>Fraxinus excelsior</i>									
<i>Quercus petraea</i>									
<i>Pinus sylvestris</i>	2		2	48	7.1	2	1-year-old seedlings from nursery	Limed	Crookshanks <i>et al.</i> 1998
<i>Nothofagus fusca</i>									
<i>Pinus radiata</i>	2		8	31	17.0	1 [†]	Beech seedlings from forest; clonal tissue culture pines	Native beech forest soil added; irrigated and fertilized	Hogan <i>et al.</i> 1997

Reps, replications. * Levels of CO₂ included ambient (~350–360 p.p.m.), elevated (~600–700), and in some studies intermediate levels. [†]Exposure is continuing.

Table 2. Photosynthetic enhancement ratios (elevated/ambient, E/A) observed in field grown trees exposed to CO₂ concentrations ≈250–350 p.p.m. above ambient. Ratios in column 6 were calculated from the photosynthetic rates measured at the growth concentration, and those in columns 7 and 8 from rates measured at common C_a. Values in the table represent the mean ratio for an experiment within each species and interacting treatment. Photosynthetic rates were taken from text, tables or estimated from figures in the sources cited. Trends within a treatment (seasonal, with temperature, with decreasing water potential, etc.) are discussed in the text

Species	Additional treatment	Year*	Times [†]	Reference	Photosynthetic ratio (E/A) [‡]		
					Measured at: Growth [CO ₂]	Ambient [CO ₂]	Elevated [CO ₂]
Deciduous broadleaved							
<i>Acer rubrum</i>	Shade	1st	1	Kubiske & Pregitzer 1996	1.63		
<i>Acer rubrum</i>	Sun	1st	1	Kubiske & Pregitzer 1996	1.70		
<i>Acer rubrum</i>	Ambient T.	1st–4th	11	Gunderson, unpublished	1.36		
<i>Acer rubrum</i>	Elevated T.	1st–4th	11	Gunderson, unpublished	1.51		
<i>Acer saccharum</i>	Ambient T.	1st–4th	15	Gunderson, unpublished	1.27		
<i>Acer saccharum</i>	Elevated T.	1st–4th	15	Gunderson, unpublished	1.52		
<i>Alnus glutinosa</i>		1st	4	Vogel & Curtis 1995	1.46		
<i>Betula papyrifera</i>	Sun	1st	1	Kubiske & Pregitzer 1996	1.70		
<i>Betula papyrifera</i>	Shade	1st	1	Kubiske & Pregitzer 1996	1.09		
<i>Betula pendula</i>		4th	3	Rey & Jarvis 1998	1.33	0.74	0.82
<i>Fagus sylvatica</i>		2nd	2	Epron <i>et al.</i> 1996		0.93	
<i>Liriodendron tulipifera</i> ¹		2nd–3rd	7	Gunderson <i>et al.</i> 1993	1.58		
<i>Liriodendron tulipifera</i> ²		1st	2	Wullschleger <i>et al.</i> 1992b			
		& 4th		Gunderson & Wullschleger 1994	1.62		
<i>Liriodendron tulipifera</i> ²	Coppice	4th	1	Gunderson & Wullschleger 1994	1.61	0.93	
<i>Populus grandidentata</i>		1st	8	Curtis <i>et al.</i> 1994	1.62		
<i>Populus tremuloides</i>	Low N	1st	n/a [†]	Kubiske <i>et al.</i> 1997	1.55		
<i>Populus tremuloides</i>	High N	1st	n/a [†]	Kubiske <i>et al.</i> 1997	1.98		
<i>Populus deltoides</i> × <i>P. nigra</i>	High fertility	1st	7	Curtis <i>et al.</i> 1995	1.40	0.76	0.87
<i>Populus deltoides</i> × <i>P. nigra</i>	Low fertility	1st	7	Curtis <i>et al.</i> 1995	1.40	0.77	0.90
<i>P. trichocarpa</i> × <i>P. deltoides</i>		1st	1	Ceulemans <i>et al.</i> 1997	2.64	1.14	1.18
<i>Populus deltoides</i> × <i>P. nigra</i>		1st	1	Ceulemans <i>et al.</i> 1997	2.84	1.20	1.13
<i>Populus hybrids</i> (2 clones)	Coppice	3rd	2	Will & Ceulemans 1997	1.60	0.97	0.97
<i>Quercus alba</i> ¹		2nd–3rd	7	Gunderson <i>et al.</i> 1993	1.79		
<i>Quercus alba</i> ²		1st	1	Wullschleger <i>et al.</i> 1992b	1.51		
<i>Quercus alba</i> ²		4th	3	Gunderson, unpublished		0.85	
<i>Quercus rubra</i>	Shade	1st	1	Kubiske & Pregitzer 1996	2.63		
<i>Quercus rubra</i>	Sun	1st	1	Kubiske & Pregitzer 1996	2.57		
<i>Quercus rubra</i>		2nd	5	Dixon <i>et al.</i> 1995	1.54		
Evergreen broadleaved							
<i>Citrus aurantium</i>		2nd–3rd	n/a [†]	Idso & Kimball 1991	2.22		
<i>Eucalyptus tetradonta</i>		1st–3rd	3	Eamus <i>et al.</i> 1995	1.29		
<i>Mangifera indica</i>		1st–3rd	9	Goodfellow <i>et al.</i> 1997	1.20		
<i>Mangifera indica</i>		3rd	1	Goodfellow <i>et al.</i> 1997		0.96	0.68
<i>Nothofagus fusca</i>		2nd	1	Hogan <i>et al.</i> 1997	1.45	0.87	0.68
<i>Quercus ilex</i>		1st–3rd	7	Scarascia-Mugnozza <i>et al.</i> 1996	1.69		
Conifers							
<i>Picea alba</i>		2nd	5	Dixon <i>et al.</i> 1995	1.43		
<i>Pinus ponderosa</i>		6th	1	Tissue <i>et al.</i> 1998	1.53		
<i>Pinus radiata</i>		2nd	1	Hogan <i>et al.</i> 1997	1.47	0.80	0.95
<i>Pinus sylvestris</i>	Ambient T.	4th	3	Kellomäki & Wang 1996	1.41	0.91	0.87
<i>Pinus sylvestris</i>	Elevated T.	4th	3	Kellomäki & Wang 1996	1.62	1.02	0.96
<i>Pinus taeda</i>		1st–2nd	8	Tissue, Thomas & Strain 1996	1.73		
		3rd–4th		Tissue <i>et al.</i> 1997			

*Measurements spanned these years or growing seasons after enrichment began. [†]Number of times photosynthesis was measured, i.e., number of reported values contributing to the means listed; n/a indicates more than once, exact number not stated. [‡]E/A: rate in elevated-CO₂ leaves/ rate in ambient-CO₂ leaves, measured at the concentrations indicated ^{1,2} indicate two separate experiments with the same species.

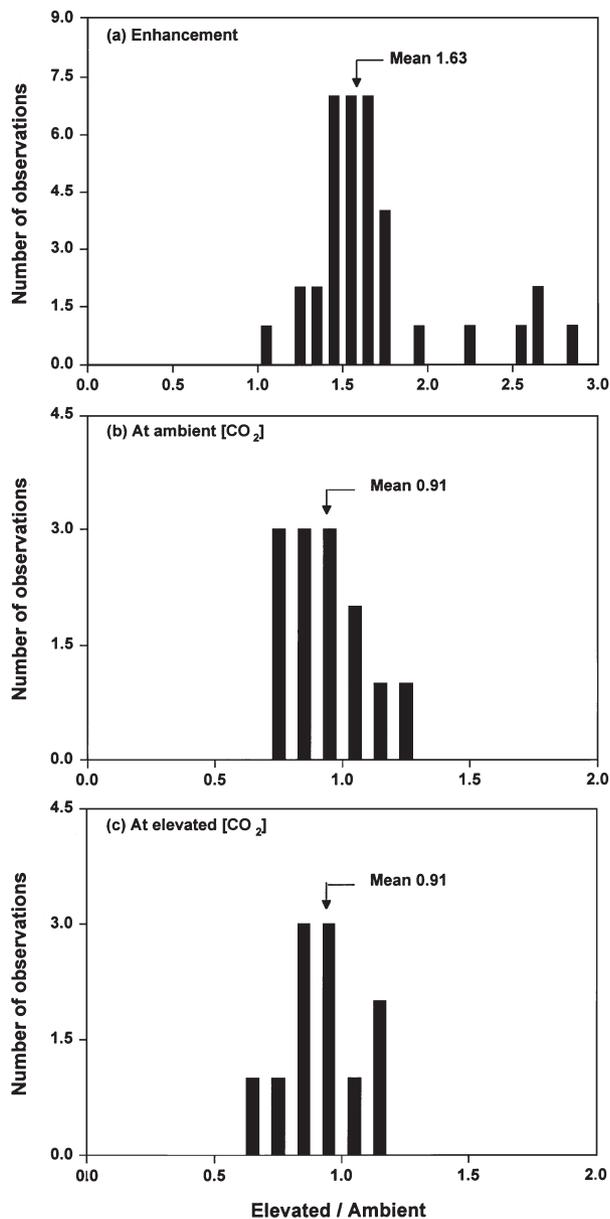


Figure 1. Frequency distribution for the relative photosynthetic responses of field-grown trees under CO₂ enrichment compared to those at ambient [CO₂]. Frequency indicates the number of observations (see Table 2) within each ratio interval for (a) leaves measured at their respective growth [CO₂], (b) leaves measured at ambient [CO₂] concentrations regardless of growth [CO₂], and (c) leaves measured at a common elevated [CO₂].

and there was no evidence of a nitrogen reallocation from ribulose biphosphate carboxylase/oxygenase (rubisco) to other photosynthetic systems (Curtis *et al.* 1995). In a fourth study (Tissue, Griffin & Ball 1998), annual soil nitrogen fertilization had no significant effect on photosynthetic parameters.

Under some circumstances, responses to CO₂ might be reduced if water deficits are severe enough to limit photosynthetic enzymatic activity, but responses are more likely

to be enhanced if elevated [CO₂] reduces the importance of drought-induced stomatal limitation (Chaves & Pereira 1992). Two experiments with mature trees, in which the response to CO₂ enrichment and changes in leaf water potential were tracked during natural droughts, support the latter hypothesis. The relative photosynthetic stimulation increased to 100% enhancement at water potentials of -4.5 MPa (Scarascia-Mugnozza *et al.* 1996), particularly at elevated temperatures (Kellomäki & Wang 1996). Enhancement was likewise greater during drought for *Picea abies* saplings in an unreplicated open-top chamber experiment, although the four *Quercus rubra* saplings in the same chamber showed the opposite response (Dixon, LeThiec & Garrec 1995). Goodfellow, Eamus & Duff (1997) reported greater impacts of CO₂ enrichment during the tropical dry season when stomatal conductance was low, and leaf water potential was maintained.

Many of the differences in CO₂ effects within studies and perhaps between studies can be explained by temperature differences. As discussed by Long (1991), the relative affinity of rubisco for CO₂ decreases markedly with increasing temperature, but elevated CO₂ concentrations increase the competitive inhibition of oxygenation such that the relative stimulation of assimilation by elevated CO₂ increases with temperature, and the temperature optimum for assimilation increases with increasing [CO₂]. Experiments with temperature manipulation treatments confirm this with higher CO₂ enhancement ratios for trees growing in temperatures raised 2–4 °C above the ambient chambers (Table 2; Kellomäki & Wang 1996; C. Gunderson, unpublished results). Idso *et al.* (1995) compared rates measured at leaf temperatures from 30 to 46 °C over four growing seasons. Relative stimulation by CO₂, already higher at these temperatures than at the more moderate conditions of many studies, increased with temperature, and sharply so, as assimilation rates in the ambient CO₂ trees approached zero at the highest temperatures. Temperature is also a factor in some seasonal patterns reported for CO₂ responses, for example, much of the difference in enhancement of assimilation in *Pinus taeda* in summer months (60–130% increase) versus winter months (14–44%), is explained by seasonal temperature differences (Tissue, Thomas & Strain 1997; Lewis, Tissue & Strain 1996).

Most of the results discussed above are from single healthy leaves at comparable leaf age and position, measured at light saturation and, in some cases, under idealized conditions, minimizing leaf-to-air vapour pressure difference and controlling temperature. This approach minimizes factors that might confound the interpretation of photosynthetic response per se, but does not address the question of canopy-level effects on assimilation, which will change with plant development. Pertinent experimental techniques include single-leaf measurements at multiple positions in the canopy, measurements of the entire canopy (which is difficult for larger trees), and light-response curves to estimate CO₂ effects within a closed canopy. Young saplings of both *Liriodendron tulipifera* and *Quercus alba* sustained

comparably higher assimilation rates at multiple canopy positions and leaf ages (Gunderson, Norby & Wullschleger 1993), but photosynthetic enhancement in 1-year-old needles of *Pinus radiata* was lower than in current needles (31% versus 64%; Turnbull *et al.* 1998). One-year-old *Populus tremuloides* demonstrated greater photosynthetic enhancement by CO₂ in the lower half of the crown, without a change in N distribution within the canopy (Kubiske *et al.* 1997), although mid-crown leaves (only) exhibited reductions in photosynthetic capacity. Measurements in these studies, however, were made at light saturation, and in young trees without much self shading. Measurements of *Pinus elliottii* seedlings incorporated self-shading effects by use of a whole-tree cuvette (Garcia *et al.* 1994), but it is essential with such techniques to separate the CO₂ effect on photosynthesis (1.9 times higher in a short-term measurement) from the combined effects of increased canopy leaf area and higher photosynthesis (2.8 times higher). A related approach involved microcosms enclosing small stands of young *Fagus sylvatica* trees, where whole ecosystem measurements were compared to single leaf measurements via modelling procedures (Overdieck 1993).

Light attenuation within a mature forest canopy and the interactions between [CO₂] and leaf acclimation to light environment are important factors in evaluating CO₂ responses at the canopy level, but these issues are not easily addressed in open-top chambers. Single-leaf measurements of light-response curves generally reveal an increase in apparent quantum yield (Kubiske & Pregitzer 1996; Goodfellow *et al.* 1997) and a decrease in light compensation point (Kubiske & Pregitzer 1996) with CO₂ enrichment, because elevated [CO₂] inhibits photorespiration (Long & Drake 1991). A higher initial slope for assimilation versus light has also been noted at the canopy level (Garcia *et al.* 1994) for seedlings at elevated [CO₂]. Variability in leaf response to CO₂ was reported in relation to the light environment and a species' shade tolerance (Kubiske & Pregitzer 1996), and with seasonal and diel variation in irradiance (Goodfellow *et al.* 1997). In general, however, higher CO₂ concentrations should enhance carbon gain at low light levels, for example, in the lower canopy, in understory plants, and on cloudy days.

As indicated by many single and multiyear studies, sustained photosynthetic responses to elevated CO₂ (Table 2, Fig. 1a) have disproved the conjecture that days, weeks or months of exposure to CO₂ would result in a loss of most of the enhancement effect. These data do not by themselves, however, indicate whether there may have been a more subtle biochemical or physiological 'acclimation' to growth at elevated CO₂, a reduction in photosynthetic capacity at equivalent conditions, or a partial loss of enhancement with time. Results of this type have been reported in trees grown in pots, and in other types of plant material (reviewed elsewhere: Gunderson & Wullschleger 1994; Sage 1994; Drake, González-Meler & Long 1997). When reduced stimulation has been found, it has been postulated to arise from either end-product inhibition (i.e. down-regulation by carbohydrate accumulation) or as a

result of what may be termed acclimation, a suite of biochemical and physiological adjustments considered to improve plant performance through increased efficiency in use of resources (Sage 1994). These internal changes could be extremely important if they were to have an impact on net assimilation such that photosynthetic stimulation by CO₂ was lost over time, or was much lower than predicted from short-term measurements.

Such major losses of enhancement have not been demonstrated for trees rooted in the ground. Nevertheless, there have been attempts to resolve smaller differences in foliage developed under CO₂ enrichment. A downward trend in photosynthetic enhancement through time might be revealed by repeated measurements during the course of an experiment. There was no such trend in *Acer saccharum*: in ambient temperatures the 25% enhancement on the first day of exposure (C. Gunderson, unpublished results) was almost the same as the 4-year mean (Table 2). Enhancement was higher (53%) in *Eucalyptus tetrodonta* after 2.5 years than in previous years (Eamus *et al.* 1995). Several studies report seasonal differences in sensitivity to CO₂, but these differences cannot be characterized as a general downward trend over time and were often attributed, as indicated above, to other environmental factors, e.g. moisture availability (Dixon *et al.* 1995; Scarascia-Mugnozza *et al.* 1996; Kellomäki & Wang 1996; Goodfellow *et al.* 1997) or temperature (Lewis *et al.* 1996), or to a seasonal change in source-sink balance (Rey & Jarvis 1998). In some cases, enhancement was greater at the end of a growing season, attributed to effects of N availability on late season dynamics (senescence), either from applied N (Curtis *et al.* 1994) or from symbiotic N₂ fixation (Vogel & Curtis 1995).

A second method of assessing photosynthetic capacity in trees from two CO₂ treatments has been 'reciprocal transfer', switching the CO₂ concentrations, either of the whole chamber (Goodfellow *et al.* 1997), or more commonly, of only the leaf cuvette. For the nine species-treatment combinations where these data are available (Table 2), the ratio of enriched-grown foliage to ambient (E/A) ranged from 0.68 to 1.15, for a geometric mean of 0.92 — only an 8% decrease in capacity (Table 2, Fig. 1b,c). This is in marked contrast with the 21% decline calculated from 20 studies of pot-grown tree seedlings (Gunderson & Wullschleger 1994) and more in agreement with the nonsignificant 1% decline noted for trees in pots larger than 0.5 dm³ (Curtis & Wang 1998) and the 7% decline for a variety of species in rooting volumes > 10 dm³ (Drake *et al.* 1997). These types of measurements are designed to represent photosynthesis at equivalent conditions, and therefore a ratio less than one purports to indicate a loss of photosynthetic capacity. However, as pointed out by Goodfellow *et al.* (1997), stomatal conductance (g_s) may remain lower in foliage grown under elevated CO₂, even at equivalent cuvette concentrations (C_a), perhaps because of reduced stomatal density (cf. Rey & Jarvis 1998). If a lower g_s reduces intercellular CO₂ concentrations (C_i) in elevated CO₂-grown foliage, as in *Mangifera indica* (Goodfellow *et al.* 1997), then E/A ratios

at a common C_a would not represent differences in biochemical capacity at equivalent conditions.

Measurement at equivalent C_i can be assured with the development of A/C_i curves, that is, net assimilation measured at multiple CO₂ concentrations for which C_i are calculated based on stomatal conductance. These curves can also be used to estimate the carboxylation efficiency [V_{cmax} , the capacity of rubisco to carboxylate ribulose biphosphate (RuBP)] and RuBP regeneration capacity mediated by electron transport (J_{max}) (Sage 1994; Lewis *et al.* 1996). Little or no difference was reported between the A/C_i curves of ambient and elevated CO₂-grown foliage in four species (*Liriodendron tulipifera* and *Quercus alba*: Gunderson *et al.* 1993; *Pinus taeda*: Ellsworth *et al.* 1995; Lewis *et al.* 1996; *Pinus sylvestris* at two temperature treatments: Kellomäki & Wang 1996). The A/C_i curves of N₂-fixing *Alnus glutinosa* were identical early in the season, but V_{cmax} was 16% higher in high-CO₂ foliage later in the season (Vogel & Curtis 1995). Reductions in the A/C_i response were seen in high-CO₂ foliage of *Populus tremuloides*, but only in the middle of three canopy positions (Kubiske *et al.* 1997). V_{cmax} was 12–20% lower in *Populus deltoides* × *P. nigra* in mid-September, but not in early August (Curtis *et al.* 1995). In contrast, elevated CO₂-grown *Betula pendula* had significantly lower A/C_i curves in August and September of the fourth year, and V_{cmax} and J_{max} were numerically lower even in June (Rey & Jarvis 1998). The reduction in V_{cmax} increased from 9% to 23% over the course of the season, which is in agreement with a consistently lower and decreasing E/A ratio at equal C_a (Table 2). An even larger reduction was seen in V_{cmax} and J_{max} (36 and 21%, respectively) of *Pinus ponderosa* in September of the sixth year of CO₂ enrichment (Tissue *et al.* 1998), although photosynthesis at the growth concentration was still stimulated 53%. In *Picea abies*, A/C_i curves were not affected in June, but in September were lower in foliage from the elevated CO₂ treatment (Marek, Kalina & Matoušková 1995). The A/C_i curves from current-year needles of *Pinus radiata* showed no differences even late in the growing season, but were lower in 1-year-old needles at that time (Turnbull *et al.* 1998).

From the range of responses obtained from A/C_i curves, (one increasing, seven no change, one decreasing only at one of three canopy positions, and five decreasing later in the season in at least some foliage), it is apparent that prolonged growth at elevated [CO₂] does not result in a consistent down-regulation of photosynthetic parameters. The pattern does suggest a potential decrease in both V_{cmax} and J_{max} , particularly late in the season, concurrent with decreases in measured rubisco content (and thus activity per unit leaf area) (Tissue *et al.* 1997, 1998; Rey & Jarvis 1998; Turnbull *et al.* 1998), although decreases in rubisco activity, measured biochemically, can occur with little effect on V_{cmax} (Lewis *et al.* 1996; Drake *et al.* 1997).

In most cases, leaf mass per unit area is higher with growth at elevated [CO₂], and, as discussed later, in many cases, leaf nitrogen concentrations decrease while starch, and, less frequently, soluble sugar concentrations increase

(cf. Körner & Miglietta 1994). These changes in tissue chemistry form the basis for proposed mechanisms of acclimation based on N reallocation and feedback-driven down-regulation (Drake *et al.* 1997), but they are not necessarily indicative of either phenomenon. In fact, in many of the studies in Table 1, these changes occur without any evidence of altered photosynthetic response, and conversely, some of the changes in A/C_i curves noted above were not associated with changes in N or sugars. With respect to the N reallocation hypothesis, Drake *et al.* (1997) point out that at higher temperatures and increasing [CO₂], a leaf can sustain a substantial loss in rubisco content (which accounts for a significant fraction of foliar N) without an effect on assimilation rate. A model of *Pinus sylvestris* trees in open-top chambers indicated that crown photosynthesis increased 22–27% in elevated CO₂ with only marginal effects of the observed adjustment in leaf biochemistry (Kellomäki & Wang 1997a). Thus, although there are some consistent changes in leaf properties with growth in elevated CO₂, many of the previously reported changes in leaf biochemistry are less pronounced in trees planted in the ground and appear to have minimal impact on photosynthetic enhancement. Seasonal changes in carbohydrate status associated with the cessation of above-ground growth and a reduction in sink strength may explain some of the observations of late-season reductions in photosynthetic response (e.g. Epron, Liozon & Mousseau 1996). Nevertheless, it is important to emphasize that changes in leaf biochemistry, including seasonal declines in V_{cmax} or rubisco, do not eliminate a photosynthetic response to elevated CO₂.

All of the evidence from field-grown trees suggests a continued, and surprisingly consistent, stimulation of photosynthesis, ≈ 60% for a 300 p.p.m. increase in [CO₂]. There is, at present, little reason to expect a long-term loss of sensitivity to CO₂ as suggested by earlier pot studies of trees. Research on the response of photosynthesis to rising CO₂ will continue, of course, to extend our understanding beyond 6-year exposures and to resolve questions about seasonal changes in photosynthetic biochemistry.

Canopy structure

The carbon uptake of a tree or a forest stand cannot be calculated simply from the rates of net photosynthesis of individual leaves. These rates must be integrated over the entire canopy and over the growing season. Tree and forest models accomplish this through calculation of the light extinction within a canopy for a given leaf area index (LAI), coupled with information on the light response of photosynthesis and seasonal trends in temperature, water, and other environmental factors that influence net carbon uptake (Kellomäki & Wang 1997a). Tree growth in elevated CO₂ has the potential to alter many of these relationships. Any effect of CO₂ on maximum LAI, the seasonal development or structure of the canopy, or the single-leaf response to gradients within the canopy will change the relationship between instantaneous net carbon uptake of individual leaves and annual carbon uptake of the whole canopy.

Although canopy structure and processes are clearly critical components of tree response to increasing atmospheric CO₂, there are very few data from CO₂ enrichment studies that are relevant to our scale of interest. Consider first the central question of whether the LAI of a forest stand will be different in a high-CO₂ world. The leaf area of the seedlings and saplings grown in open-top experiments has usually increased with CO₂ enrichment. Leaf area of *Pinus taeda* was 41% greater in elevated versus ambient CO₂ after 4 years (Tissue *et al.* 1997), and it increased 8–18% in *Populus* clones (Ceulemans, Jiang & Shao 1995). An increase in CO₂ concentration resulted in a higher leaf area via an increase in flush length and number of fascicles in *P. sylvestris* (Kellomäki & Wang 1997a). Leaf area of *Citrus aurantium* trees was increased primarily because CO₂-enriched trees had 78% more leaves than trees in ambient CO₂, but average leaf size also increased by 13% (Idso, Kimball & Hendrix 1993a). The increase in leaf area of *Quercus alba* saplings in elevated CO₂ (Fig. 2) also can be attributed to increased leaf number; leaf size and shape changed little (Gregory 1996).

These observations of increased leaf area in elevated CO₂ do not indicate a specific stimulatory effect of CO₂ on leaf production. In the *Q. alba* experiment, for example, leaf area increased with CO₂ enrichment less than whole-plant mass did; hence leaf area ratio (LAR) was lower in elevated CO₂ (Fig. 2). LAR was reduced in *Pinus taeda* as well (Tissue *et al.* 1997). In a compilation of all CO₂ experiments with trees (including growth chamber experiments with seedlings in pots), LAR was on average 15% less in elevated CO₂ (Wullschleger *et al.* 1997a). Hence, we can conclude that the data from open-top chambers mostly show that larger plants had more leaf area.

Unfortunately, these observations tell us little about the potential CO₂ effect on LAI in a closed-canopy forest where LAI is constrained by nutrients, water or light. There have been no manipulative studies in which the experimental trees grew long enough to maintain a closed canopy for several years. Elevated CO₂ might be expected to increase LAI if the light-compensation point for photosynthesis is higher such that leaves are retained deeper in the canopy. Alternatively, if elevated CO₂ exacerbates nutrient constraints, LAI could be reduced. The observation that LAR is reduced in CO₂-enriched trees might also suggest that LAI will be reduced. The only direct measure of a CO₂ effect on LAI comes from unreplicated observations of two coppice forests near CO₂ vents in Italy, where the trees have been exposed to elevated CO₂ concentrations throughout their 35–40 years. There was no difference in LAI between the CO₂-enriched sites and nearby control sites, although LAR was lower in the CO₂-enriched sites (Hättenschwiler *et al.* 1997b).

Changes in canopy architecture could be important even if LAI is not changed, especially if the photosynthetic responses to CO₂ change with light or canopy position. Arnone & Körner (1993) suggested that changes in the vertical leaf display and crown structure might alter the red/far red ratio of light reaching understory tree seedlings,

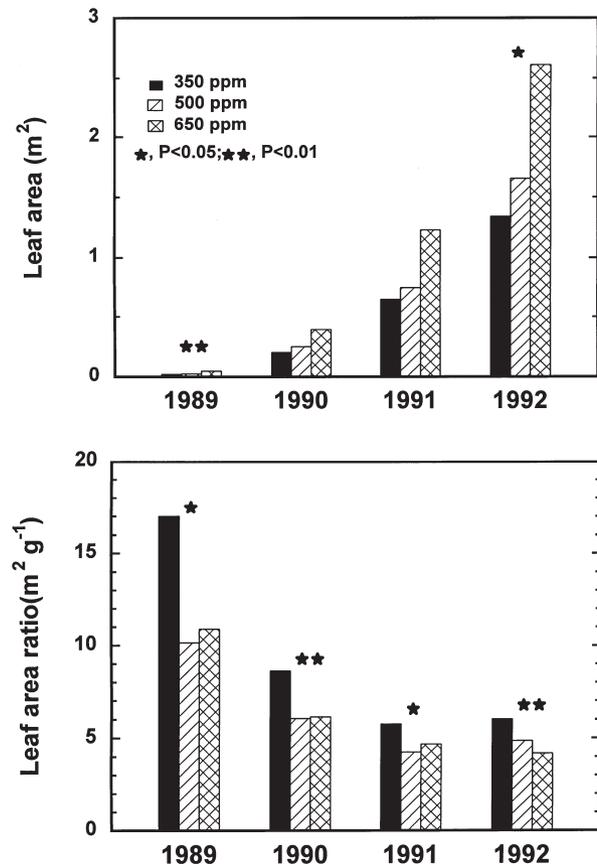


Figure 2. Leaf area and leaf area ratio (leaf area divided by above-ground plant dry mass) of *Quercus alba* trees grown in ambient and elevated CO₂ (Norby *et al.* 1995). The plants were grown in open-top chambers with two replicates for each of the three CO₂ concentrations from April 1989 until September 1992. Leaf area was determined from collections made at the end of each growing season as the leaves abscised. Above-ground plant dry mass was estimated from height and diameter measurements in 1989–91 and was measured directly when the plants were harvested in September 1992.

thereby affecting their pattern of growth. *Pinus sylvestris* trees not only had more leaf area in elevated CO₂, but there was also a shift in foliage distribution with relatively more leaves toward the base of the crown in CO₂-enriched trees (Kellomäki & Wang 1997a). These adjustments might be important for maximizing light harvesting and minimizing self-shading (Kellomäki & Wang 1997a). Increased secondary branching in elevated CO₂ was indicated by Idso, Kimball & Allen (1991) and Ceulemans *et al.* (1995). Norby *et al.* (1996), however, saw no change in any index of canopy structure in *Quercus alba* or *Liriodendron tulipifera*. Increasing our understanding of branch morphology and crown characteristics will aid in efforts to scale results of physiological studies to the tree or stand level, as large-scale canopy function is an integration of physiological processes and structure at smaller scales.

Recent observations of large-scale carbon fluxes by the eddy covariance approach have demonstrated that canopy

phenology, the duration of leaf display, is an important determinant of year-to-year variation in annual net carbon flux (Goulden *et al.* 1996). The possibility of changes in phenology in response to elevated CO₂ was an important reason to conduct experiments in the field over several growing seasons. Most of the observations of phenology in these studies, however, have been somewhat casual, and it is difficult to determine if there are any general patterns. Gunderson *et al.* (1993) quantified the timing of fall senescence in *Liriodendron tulipifera* and *Quercus alba* by measuring the decline in chlorophyll content and the time course of leaf abscission. There were no effects of elevated CO₂ in either species. One clone of *Populus* exhibited delayed bud burst in elevated CO₂, whereas another clone exhibited advanced bud set (Ceulemans *et al.* 1995). *Picea sitchensis* and *Castanea sativa*, growing in pots in field chambers, exhibited both delayed bud burst and advanced bud set (El Kohen *et al.* 1993; Murray *et al.* 1994), but there were no effects of CO₂ on the bud phenology of four other tree species (Murray & Ceulemans 1998). Elevated temperature accelerated bud burst in *Pseudotsuga menziesii*, but elevated CO₂ counteracted this effect; elevated CO₂ also decreased bud hardiness during cold hardening and dehardening (Guak *et al.* 1998). Increased temperature had important effects on the timing of spring bud break and autumn leaf senescence in *Acer saccharum* and *A. rubrum*, but there were no important or consistent effects of elevated CO₂ (Norby *et al.* 1998). There is at yet no basis for ascribing this variation in phenological response to increased CO₂ to inherent differences between species in their ability to optimize the timing of developmental events. Nevertheless, competitiveness and survival of trees can depend on the ability to avoid having periods of growth coincide with periods of subzero temperatures, and a differential response to elevated CO₂ could alter competitive relationships and stand structure.

Respiration

The supposition that trees will maintain higher rates of leaf and canopy photosynthesis when grown at elevated CO₂ appears to be supported by many field experiments. Photosynthesis is, however, only one determinant of a tree's carbon balance, and researchers have in recent years expanded their focus to consider also the respiratory loss of carbon by woody plants exposed to atmospheric CO₂ enrichment. These studies have provided periodic estimates of respiration for both seedlings and saplings grown at ambient and elevated CO₂ (Idso & Kimball 1992a; Wullschlegel, Norby & Hendrix 1992b; Vogel & Curtis 1995; Curtis *et al.* 1995; Ceulemans *et al.* 1997) and have attempted to identify the sensitivity of growth and maintenance respiration to elevated CO₂ in leaves (Wullschlegel & Norby 1992; Wullschlegel, Norby & Gunderson 1992a; Will & Ceulemans 1997) and stems (Wullschlegel, Norby & Hanson 1995b; Carey, DeLucia & Ball 1996; Dvorak & Oplustilova 1997). The energetic costs of tissue construction have similarly been examined in leaves, stems, and

roots for field-grown trees exposed to elevated CO₂ (Carey *et al.* 1996; Wullschlegel *et al.* 1997b), and these effects have, in potted *Pinus ponderosa* and *P. taeda* seedlings, been attributed to CO₂-induced changes in the biochemical composition of leaves (Griffin, Winner & Strain 1996b).

While these studies have advanced to some extent our understanding of the potential response of woody plant respiration to CO₂ enrichment, it is unfortunate that no strong scientific consensus has yet emerged from these observations. Single-leaf rates of respiration are often reported to be lower for field-grown trees exposed to elevated CO₂ (Idso & Kimball 1992a; Wullschlegel *et al.* 1992a; Teskey 1995; Ceulemans *et al.* 1997). These effects range from a 14% suppression of respiration for needles of *Pinus taeda* in branch bags (Teskey 1995) to 60% or more for one clone of hybrid poplar (Ceulemans *et al.* 1997). There are, however, equally compelling observations that respiration is unresponsive to CO₂ enrichment (Vogel & Curtis 1995; Curtis *et al.* 1995; Ceulemans *et al.* 1997; Will & Ceulemans 1997). This inconsistency of response has been observed both within individual experiments and between studies conducted by different investigators. Ceulemans *et al.* (1997), for example, studied the respiratory response of two contrasting *Populus* hybrids grown at ambient and elevated CO₂ in open-top field chambers. Elevated CO₂ had no long-term effect on leaf respiration for the slow-growing clone Robusta (*P. deltoides* × *P. nigra*), but rates of respiration for the fast-growing clone Beaupré (*P. trichocarpa* × *P. deltoides*) were more than 60% lower at elevated CO₂ concentrations. Genotypic variation such as this, if substantiated, could be used to explore mechanisms whereby respiration changes in response to CO₂ enrichment. Unfortunately, these clonal differences were not observed in a subsequent study conducted on coppice regrowth of the original plant material (Will & Ceulemans 1997), so there is some question as to whether the clone-specific response observed by Ceulemans *et al.* (1997) represents true genetic variation or instead reflects variability attributable to experimental protocol.

There are, of course, other possibilities that could be invoked to explain the highly variable and inconsistent response of respiration to CO₂ enrichment: complications caused by expressing respiration on a leaf mass or area basis, stages of plant development, leaf age and carbohydrate composition, chamber leaks and artifacts resulting from methodology, and interacting factors such as temperature or nutrient status of the measured tissues. These confounding factors have seldom been considered in measurements of leaf respiration at either ambient or elevated CO₂, and such uncertainties are currently hindering progress in this area. Steps must be taken to resolve these issues by conducting field-based studies that systematically address the short-term direct effects and long-term acclimation effects of elevated CO₂ on leaf respiration. A direct effect is defined here as an immediate response in which rates of respiration are altered by a change in CO₂ surrounding a leaf or whole plant; it is a reversible effect and occurs within minutes of a step change in CO₂ (Drake *et al.* 1999).

An acclimation effect, by comparison, occurs when rates of respiration for trees grown in elevated CO₂ differ from those grown in ambient CO₂, with the stipulation that all measurements are made at a common CO₂ partial pressure. This latter definition implies that the acclimation effect is persistent and thus reflects an intrinsic change in tissue chemistry (e.g. N or protein content) or in some whole-plant process (e.g. growth or biomass allocation) that is subsequently reflected in measurements of respiration.

The utility of separating direct from acclimation effects has been nicely demonstrated in the branch-bag studies of Teskey (1995) and in the whole-shoot investigations of Griffin, Ball & Strain (1996a). Each of these studies observed that short-term increases in CO₂ could elicit an immediate and apparently reversible suppression of respiration. This direct effect ranged from a 6–14% suppression of respiration as [CO₂] surrounding branches of 21-year-old *Pinus taeda* was raised from ambient to ambient + 330 p.p.m. (Teskey 1995) and from a 3–13% inhibition of respiration as [CO₂] was increased from 350 to 700 p.p.m. around whole-shoots of *Pinus ponderosa* seedlings (Griffin *et al.* 1996a). Although this latter study was conducted on potted seedlings, it nonetheless illustrates an experimental approach whereby the direct and acclimation effects of elevated CO₂ can be separately addressed. This is an important consideration, as Griffin *et al.* (1996a) demonstrated that the magnitude of a direct suppression of needle respiration was correlated in *P. ponderosa* with longer-term changes in tissue C/N ratios; the direct effect of elevated CO₂ on respiration was greatest in shoots with a higher C/N ratio. These findings are particularly relevant given the often reported observation that leaf [N] is lower in woody plants exposed to long-term CO₂ enrichment (Curtis & Wang 1998). Thus, barring unforeseen changes in leaf carbon content, a decrease in tissue C/N ratios may strengthen any direct response of leaf respiration to elevated CO₂ concentration.

A mechanistic explanation and a series of testable hypotheses are urgently needed for the direct and, to a lesser extent, the acclimation effects of elevated CO₂ on respiration. It is likely that without such an explanation future measurements of leaf respiration at ambient and elevated CO₂ will be viewed cautiously. Once a cause-and-effect relationship is proposed, however, there will still be a critical need to integrate this information within the context of whole-tree responses to CO₂ enrichment. Wang, Rey & Jarvis (1998) conducted such a prototype analysis for young *Betula pendula* trees and not only considered the effects of elevated CO₂ on biomass growth, but integrated this information with known or suspected effects of atmospheric CO₂ on photosynthesis and tissue-specific rates of respiration. Trees in their fourth year of growth at elevated CO₂ were 48% larger than those grown at ambient CO₂, and during the growing season trees in the ambient and elevated CO₂ treatments increased their biomass by 4–5-fold. The annual loss of carbon (g C tree⁻¹ year⁻¹) for all plant tissues combined (leaves, stems, and roots) was about equally divided between growth (45%) and maintenance

(55%) respiration, and accounted for 31–38% of the total CO₂ assimilated in gross photosynthesis (Fig. 3). One surprising finding from this analysis was that a 23% reduction in leaf respiration in elevated CO₂ had little impact on the overall carbon budget of these rapidly growing trees (Wang *et al.* 1998). However, if Wang *et al.* (1998) assumed that both growth and maintenance respiration were reduced in elevated CO₂, then CO₂-enriched trees were simulated to produce and maintain ≈ 60% more leaf biomass (and 43% more leaf area) per tree with an additional respiratory cost of less than 10% (332 versus 361 g C tree⁻¹ year⁻¹). A similar conclusion was reached by Norby *et al.* (unpublished results) in their carbon budget analysis of *Quercus alba* where CO₂-induced reductions in growth and maintenance respiration enabled trees at elevated CO₂ to produce and maintain throughout the season more than 90% more leaf biomass at an additional respiratory cost of less than 15% (160 versus 181 g C tree⁻¹ year⁻¹). These analyses suggest that while the effects of elevated CO₂ on leaf growth and maintenance respiration may play only a limited role in whole-plant carbon budgets, these effects could nonetheless be of some 'local' significance to the carbon balance of tree canopies.

The carbon budget analysis of Wang *et al.* (1998) admittedly lacks explicit treatment of root turnover and the energy costs of carbohydrate translocation and nutrient uptake, although these issues are critical unknowns for the carbon balance of CO₂-enriched trees. Wang *et al.* (1998) emphasized that much uncertainty surrounds the large respiratory losses associated with fine-root production and growth of

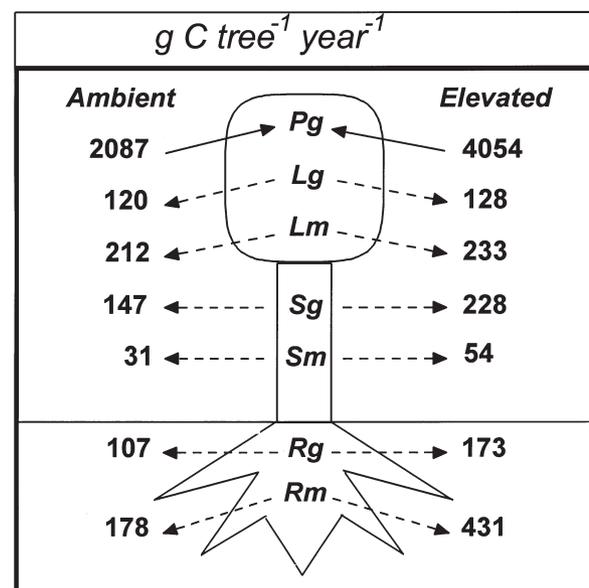


Figure 3. Annual carbon fluxes (g C tree⁻¹ year⁻¹) for young birch trees during their fourth year of growth at ambient and elevated CO₂ concentration. P_g, gross photosynthesis; L, S and R designate leaves, woody stems and roots; the subscripts _g and _m designate either growth or maintenance respiration. Data were adapted from Table 6 of Wang *et al.* (1998) with the permission of Y.-P. Wang.

the root-associated mycorrhiza at ambient and elevated CO₂ conditions. These topics have received little attention in field-grown trees. Pregitzer *et al.* (1995) suggested that the respiratory costs associated with fine-root turnover (growth and maintenance costs) may account for at least a portion of the carbon that is otherwise missing from comparisons of rates of photosynthesis and estimates of net assimilation made by destructively harvesting plants. At a more refined scale, there was a small but significant reduction in specific respiration rates of fine roots of *Fraxinus excelsior*, *Quercus petraea*, and *Pinus sylvestris* in elevated CO₂ (Crookshanks, Taylor & Broadmeadow 1998). Further uncertainty surrounds the respiratory costs of nutrient uptake in trees exposed to elevated CO₂ conditions. This point was emphasized by BassiriRad *et al.* (1996), who reported that the differential response of root uptake kinetics for NH₄⁺ and NO₃⁻ in field-grown *Pinus taeda* may have important implications for the energy requirements of nutrient acquisition by future forests. Finally, respiration is more than a process whereby carbon is lost from terrestrial vegetation; it provides carbon skeletons and energy for biosynthesis and maintenance of existing biomass, and contributes fundamentally to plant vigour. Studies that focus on the potential effects of elevated CO₂ on respiration must therefore consider also the significance of respiration for forest health and productivity.

Above-ground growth

Above-ground growth is perhaps the most obvious manifestation of the effect of CO₂ on trees in many experiments.

It would also appear to be the most important and relevant measure for projecting the response of forests to global change, for it is through growth and standing biomass that the health and functioning of a forest ecosystem is first evaluated. Above-ground growth is relatively easy to measure in comparison to root growth or the more subtle changes in gas exchange or biochemical constituents. Nevertheless, there has been a wide range of responses of tree growth reported from field experiments (Table 3), and a great deal of uncertainty on how to apply the results to the larger questions at hand.

The variety of results was apparent from the first two reports from field experiments on tree growth response to elevated CO₂. *Citrus aurantium* trees were reported to have more than doubled in size in response to CO₂ enrichment (Idso & Kimball 1992a), and that size advantage has continued for 7 years (Idso & Kimball 1997). But *Liriodendron tulipifera* trees, grown for 2.5 growing seasons in elevated CO₂, had only 27% more dry mass than trees grown in ambient CO₂, an increment that was not statistically significant (Norby *et al.* 1992). Subsequent reports have shown intermediate responses. Additional experiments in which there was no significant growth response to CO₂ are known to exist but have not been published in detail (Karnosky *et al.* 1998; D. Olszyk, personal communication). This wide range in response immediately gives rise to numerous questions: Why do the results vary? What is the 'average' response? Is there any meaning to an 'average' response? And perhaps most important, what are the implications of these results for forest response?

Species and interacting treatment	Growing seasons (no.)	E/A of above-ground woody dry mass	Reference
<i>Acer saccharum/A. rubrum</i>	4		Norby <i>et al.</i> 1997,1998
Ambient temperature		1.10	
Elevated temperature		1.73	
<i>Betula pendula</i>	4	1.55	Rey & Jarvis 1997
<i>Citrus aurantium</i>	8	2.17	Idso & Kimball 1997
<i>Fagus sylvatica</i>	2	1.91	Mousseau <i>et al.</i> 1996
<i>Fagus sylvatica/Picea abies</i>	1		Egli & Körner 1997
Low N deposition		0.99	
High N deposition		1.13	
<i>Liriodendron tulipifera</i>	2.5	1.22	Norby <i>et al.</i> 1992
<i>Pinus eldarica</i>	2	3.90	Idso & Kimball 1994
<i>Pinus ponderosa</i>	3		Walker <i>et al.</i> 1997
Low N		1.73	
Medium N		1.54	
High N		1.71	
<i>Pinus taeda</i>	4	1.90	Tissue <i>et al.</i> 1997
<i>Populus deltoides</i> × <i>P. nigra</i>	1		Pregitzer <i>et al.</i> 1995
Low N		1.19	
High N		1.45	
<i>Populus</i> hybrids	2		Ceulemans <i>et al.</i> 1996
<i>P. deltoides</i> × <i>P. nigra</i>		1.44	
<i>P. trichocarpa</i> × <i>P. deltoides</i>		1.73	
<i>Populus grandidentata</i>	1	1.06	Zak <i>et al.</i> 1993
<i>Quercus alba</i>	4	2.52	Norby <i>et al.</i> 1995

Table 3. CO₂ enrichment ratio (E/A) of above-ground dry mass of trees grown in elevated CO₂ compared to trees grown in ambient CO₂ in field experiments

The simple arithmetic mean of the enrichment response for above-ground woody dry mass of the experiments in Table 3 is 1.73, the log-adjusted mean is 1.64, and the median value is 1.55. These values are higher than but still within the range of values from previous data compilations, which were dominated by seedling studies: 1.40 (Eamus & Jarvis 1989), 1.38 for conifers and 1.68 for broadleaved trees (Ceulemans & Mousseau 1994), 1.40 (Poorter, Roumet & Campbell 1996), 1.30 (Wullschleger *et al.* 1997a), and 1.29 (Curtis & Wang 1998). Although the summary presented here ignores the important principles of meta-analysis (Curtis & Wang 1998), no degree of sophistication in calculating a mean value will circumvent the dubious value of a mean over such a wide range for understanding the response or predicting future responses. These are our most important challenges. Can the diversity of results be explained by the growth rate or growth potential of the different species, effects of environmental interactions, or differences in experimental protocol? Is there a better expression of growth that would be more informative and useful for longer-term predictions?

One of the most commonly invoked explanations for the differences in response illustrated in Table 3 (as well as for differences in photosynthesis, allocation, or almost any other measured response to elevated CO₂) is that species respond differently. On the surface this statement is almost a truism — several different species have been tested and their responses to CO₂ are different — but the conclusion is not supported by rigorous analysis. Clearly, the potential effect of species is completely confounded by many other factors, including soil conditions, weather, length of growing season, duration of the experiment, plant culture, chamber conditions and biases (which we hope do not exist!) of the experimenter. Although variation between species under identical site conditions (*Liriodendron tulipifera* versus *Quercus alba*) is large, so too is the variation within a species attributable to environmental factors (N or temperature) and the variation within a genus (*Pinus*, *Populus*) in different studies. A coherent description of differential responses to CO₂ enrichment, based on species characteristics or functional groupings of species, could be a useful input for ecosystem models, and several such schemes have been proposed (Poorter *et al.* 1996). However, without a rigorous demonstration that species characteristics were responsible for differences in the observed CO₂ response in a controlled experiment, this common reliance on 'species differences' to account for disparate responses should be avoided.

Increases in atmospheric CO₂ will be accompanied by changes in temperature, precipitation, N deposition, and tropospheric ozone. Any of these factors can be expected to modify the response of trees to CO₂, and likewise, elevated CO₂ could exacerbate or ameliorate the responses to the other factors. Some of the experiments in Table 1 have addressed these critical questions. There was no effect of elevated CO₂ on stem mass of *Populus tremuloides* grown in twice-ambient ozone, which imposed a significant stress (Karnosky *et al.* 1998). Elevated CO₂ compensated for the negative effects of increased temperature in *Acer saccharum* and

A. rubrum (Norby *et al.* 1998). There were no CO₂-temperature interactions in *Psuedotsuga menziesii* (D. Olszyk, personal communication). Interactions between CO₂ and N additions varied between experiments (Table 3), but it is questionable whether these results are a good model for interactions with deposition of N from the atmosphere (Norby 1998). These data sets from field experiments on interactions between CO₂ and other global change factors are too limited to allow general conclusions to be drawn, but this is clearly a research area that needs to be pursued. Responses to temperature increases in particular have many points of intersection with CO₂ responses and this interaction deserves more attention in future studies (Ceulemans 1997).

The largest difficulty in interpreting the data in Table 3, and a probable cause of the wide range of values, is the dominant effect of tree developmental patterns (ontogeny) on the attainment of dry matter. Tree and forest stand development must be a primary consideration in the interpretation of field experimental results and their application to longer-term predictions. In all of the experiments represented in Table 3, the trees were undergoing exponential growth for all or most of the exposure period. Larger plants have more leaf area, which increases their capacity to take up CO₂ and make more stem and leaf tissue, which further increases their capacity to take up CO₂ and grow. The effect of any factor that increases leaf area early in an experiment, such as random variation between individuals, differences in how seedlings were raised or planted, or specific effects of CO₂ enrichment, will be magnified over time by the principle of compound interest (Ceulemans & Mousseau 1994; Norby *et al.* 1996). As long as there are no constraints on leaf area production, spectacularly large CO₂ responses can occur. But in a forest stand there are always constraints to leaf area development — depending on the site, the constraint may be low nutrient availability, dry conditions, or ultimately not enough light to support the deepest leaves of a dense canopy. A CO₂ stimulation that depends on an ever-increasing leaf area index cannot be expected to be sustained, and projections that ignore this critical determinant of tree growth (Idso 1991) are certain to be false or misleading.

The large increase in final dry mass of *Quercus alba* (Norby *et al.* 1995) was shown to be a result of an early stimulation by CO₂; subsequent responses to elevated CO₂ included photosynthetic enhancement compensated by a downward adjustment in leaf area development from the expected exponential increase. The net result was a large difference in final dry mass without any increase in relative growth rate (RGR) over the last 3 years of the 4-year study. One interpretation of the growth trends in that experiment was that trees in elevated CO₂ would reach canopy closure 1 year earlier than those in ambient CO₂ (accelerated ontogeny), and at that point the relative CO₂ effect would decline. But as in other experiments, the trees were harvested while they were still in an exponential growth phase, so the projections about future responses are only speculation. Ultimately, we are interested in absolute growth rate, not relative growth, and RGR (a difficult term to apply to trees in which much of the biomass is dead) is

useful only to the extent that it guides long-term predictions from experimental data.

Leaf area constraints have probably come into play in some of the longer open-top chamber studies. The group of *Pinus ponderosa* trees in chambers had a closed canopy in the sixth and final year of the experiment, and the final increase in above-ground growth was less than that shown in Table 3 (J.T. Ball, personal communication). *Citrus aurantium* trees were grown individually, so there was not mutual shading by adjacent trees, but leaf area development was nevertheless constrained by the walls of the chamber, and the relative enhancement of above-ground growth (including fruit rinds) began to decline steadily in the third year of exposure (Idso & Kimball 1997). A decline in growth response with time, as has been observed in these experiments as well as in experiments (Bazzaz, Miao & Wayne 1993) with potted tree seedlings (where the constraint is on root development), is frequently cited as evidence that CO₂ fertilization is transitory and not likely to have a long-term influence on forest productivity. Actually, however, a decline in relative enhancement of woody biomass is expected and consistent with the patterns of tree development. Long-term predictions should not be based on the biomass enrichment ratio at the end of an experiment of only several years' duration.

If the biomass enrichment ratio is not an appropriate parameter on which to base long-term predictions, is there another expression of growth that accounts for developmental patterns and could be more robust? Norby (1996) proposed a 'canopy productivity index' (CPI) to normalize growth responses to equal leaf area. It is calculated as the

annual increment in stem mass per unit leaf area. A better expression might include woody root increment as well, but such data are rare, and an index is useful only if there are data to support it. The CPI was used by R. H. Waring (Waring & Schlesinger 1985) as 'growth efficiency' (although the term does not properly meet the definition of an efficiency), as an indication of a tree's responses to environmental stresses. The CPI is relevant only on an annual time step. It should not be confused with net assimilation rate (NAR), an instantaneous expression of growth that can be integrated over time under certain conditions. NAR has been a useful analytical tool in short-term CO₂ enrichment experiments (Norby & O'Neill 1989, 1991), but there rarely are sufficient data to support its use in longer-term experiments.

Considering all of the field experiments with broadleaf trees for which growth increment and leaf area data were available, the effect of CO₂ on CPI varied over a much smaller range than the CO₂ effect on final dry mass (Norby 1996). The average of the eight values was a 29 ± 7% enhancement (range 19–37%). We can extend this analysis to include several new studies, which slightly lowers the mean value and expands the range of observed values (Table 4). Nevertheless, the increase in CPI is still seen to be a consistent response of trees to elevated CO₂. *Pinus taeda* is the only conifer included in Table 4. Calculating a CPI for a tree with several cohorts of leaves contributing to annual stem growth, and each cohort contributing to 2 or more years of stem growth, is computationally difficult while the leaf area is still increasing. Tissue *et al.* (1997) were able to calculate the CPI in their study because of their extensive data set on leaf area.

Species	% increase in CPI	Reference
<i>Acer saccharum</i> / <i>A. rubrum</i>		Norby <i>et al.</i> 1997, 1998
Ambient temperature	11	
Elevated temperature	28	
<i>Betula pendula</i>	9	Rey & Jarvis 1997 Wang <i>et al.</i> 1998
<i>Citrus aurantium</i>	33	Idso & Kimball 1993, Idso <i>et al.</i> 1993c
<i>Fagus sylvatica</i>	31	Mousseau <i>et al.</i> 1996
<i>Liriodendron tulipifera</i>	35	Norby <i>et al.</i> 1992, 1996
<i>Populus deltoides</i> x <i>P. nigra</i> (Eugenei)		Curtis <i>et al.</i> 1995
Low fertility	22	Pregitzer <i>et al.</i> 1995
High fertility	18	
<i>Populus deltoides</i> x <i>P. nigra</i> (Robusta)	37	Ceulemans <i>et al.</i> 1995, 1996
<i>Populus trichocarpa</i> x <i>P. deltoides</i> (Beaupré)	22	Ceulemans <i>et al.</i> 1995, 1996
<i>Pinus taeda</i>	27	Tissue <i>et al.</i> 1997
<i>Quercus alba</i>	37	Norby <i>et al.</i> 1995, 1996
Average ± SD	26 ± 10	

Table 4. Response of annual stem production per unit leaf area (canopy productivity index, CPI) of field-grown trees to elevated CO₂. Table modified from table in Norby (1996)

In each experiment the trees were planted directly in the ground and exposed in open-top chambers to CO₂ partial pressures ≈350 p.p.m. (ambient) and 650–700 p.p.m. CPIs of *L. tulipifera* and *Q. alba* were calculated by regression analysis of annual stem mass increment versus leaf area. Other calculations were based on published values of mean stem dry mass or dry mass increment (or a surrogate measure) and leaf area or relative increase in leaf area.

The value of this index is that it provides a simple, measurable CO₂ response parameter from experimental studies that might be independent of tree and stand development. Badeck *et al.* (1997) criticized its use because the CPI could be highly sensitive to differences in LAI between ambient and elevated treatments. As LAI increases, the fraction of less productive shade leaves increases, and therefore CPI should decrease even while productivity per unit ground area might still increase (Badeck *et al.* 1997). The CPI declines with age and in response to environmental stress (Waring & Schlesinger 1985); hence, its absolute value at the end of an experiment should not be extrapolated into the future. But there is no obvious reason to assume that its relative response to CO₂ will change as LAI increases, although this is clearly a conjecture that must be tested. The index is also useful because it separates structural responses to elevated CO₂, such as changes in canopy structure discussed in the previous section, from functional responses — the physiological reactions of photosynthesis, respiration, carbon allocation, and so on. Structural and functional responses can be considered separately in ecosystem or global models (Woodward, Smith & Emanuel 1995), and separating them experimentally can help to focus research toward meaningful, testable hypotheses about tree response to elevated CO₂. The observation that the CPI response to CO₂ is remarkably similar across so many very different experiments under different conditions improves the prospects for success in projecting future response to atmospheric CO₂ enrichment and belies the general statement that ‘species differ in their response to CO₂’.

This analysis emphasizes the point that short-term tree growth responses cannot be extrapolated outside of the context of stand development. The very large growth responses observed in some experiments are unlikely to be sustained for many years under forest conditions. Much of the variation among experimental results can be explained by differences in leaf area development. On the basis of an analysis of growth per unit leaf area, the predicted long-term response to CO₂ (in the absence of interacting factors and environmental feedbacks) is only slightly less than that indicated by seedlings experiments: an increase of about 27% with a 300 p.p.m. increase in [CO₂]. This analysis gives rise to several questions. Is the short-term stimulation of leaf area development and tree growth an experimental artifact or an indication of an important effect of CO₂ on seedling establishment? Is the enhancement of growth per unit leaf area (or LAI at the stand scale) a robust response; that is, will this response persist after canopy closure? Alternatively, will the response to CO₂ continue to decline such that there ultimately is no difference in annual increment, and the only effect of CO₂ is the gain from the initial stimulation of growth increment? Or, will there be no gain from CO₂ at all in the end, the only effect being to shorten by several years the time over which maximum biomass is attained? These questions cannot be answered from the current database of open-top chamber experiments. Nevertheless, the observations from those

experiments have enabled us to ask better questions, and they should be an important guide to interpreting long-term data sets as they become available.

Although decades-long records of response cannot yet come from any manipulative experiments, the vegetation growing in the vicinity of the surface vents of deep geothermal springs, such as those in central Italy (Miglietta *et al.* 1993), can be a useful alternative source of data on long-term responses of trees to an atmosphere enriched in CO₂. Naturally elevated CO₂ concentrations can be assumed to have occurred for hundreds of years in these areas, and the vegetation has been subject to a concentration gradient determined by distance from the vent (Miglietta *et al.* 1993). But the CO₂ springs are not ideal experimental systems (Amthor 1995) — the exposure history and dynamics are uncertain, there are no true controls, and environmental conditions may be atypical — and the data must be interpreted with caution. Hättenschwiler *et al.* (1997a) described the tree ring record of *Quercus ilex* trees at two natural CO₂ springs in Italy. The trees have been exposed continuously to high CO₂ since they were seedlings (31–36 years), and throughout that time they have been larger than equal-aged trees in adjacent sites away from the CO₂ emissions. An analysis of the relative difference in tree ring width, however, indicated that the response to CO₂ was declining with time and had disappeared by the time the trees were 25–30 years old (Hättenschwiler *et al.* 1997a). Stem basal area of trees in elevated and ambient CO₂ was reconstructed from the tree ring records, and we can analyse this record with the assumptions that basal area is a good correlate of above-ground biomass, that the relationship between basal area and biomass is the same for trees in ambient and CO₂-enriched trees, and that the relationship has been constant through time. Figure 4(a) shows the relative CO₂ stimulation of basal area as a function of tree age at the Rapolano site, and there clearly was a steep decline in response from year 3 to year 13, but the record then levelled off at about 1.26, or a 26% increase in basal area in elevated CO₂. Annual basal area increment (Fig. 4a), which is presented as a 3-year running average to smooth out large year-to-year fluctuations, was always higher in the CO₂-enriched site, except for the last several years. Starting at year 9, the slope of BAI versus age was not significantly different from zero and centered on an enrichment ratio of 1.19. The record from Laiatico (not shown) was similar except for a sharp rise in BAI only in the control site in 4 of the last 5 years. The BAI record at Rapolano is consistent with predictions from the open-top chamber experiments. The approximate doubling of growth during the earliest years was not sustained, possibly declining as LAI reached maximum values for the sites. (There is no record of leaf area development for these stands, but it is reasonable to assume that as a coppice stand, they reached their maximum LAI fairly early; S. Hättenschwiler, personal communication). Since LAI was the same at enriched and control sites (4.0 for Rapolano and 3.5 for Laiatico; Hättenschwiler *et al.* 1997a), the data support the premise that enhancement of

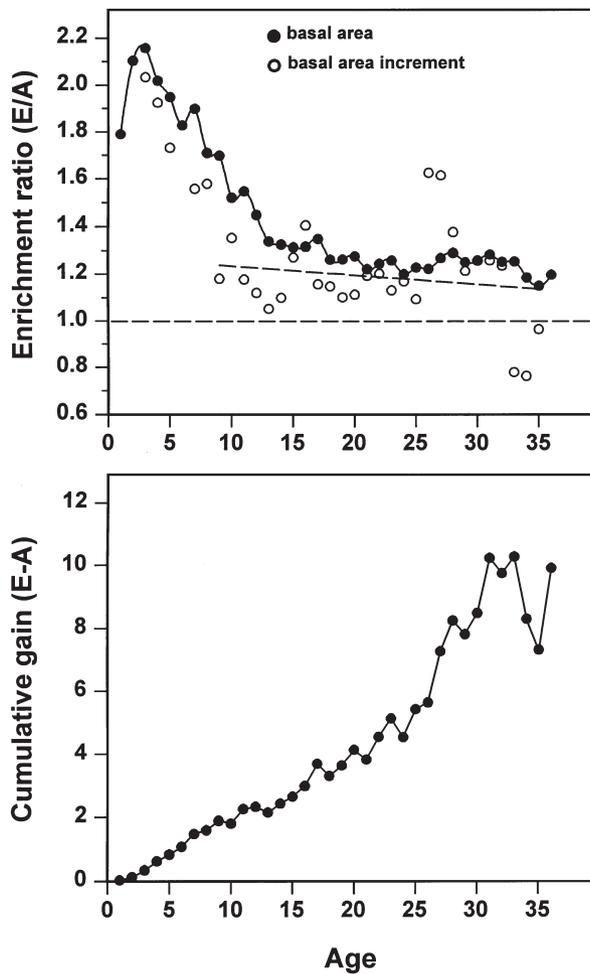


Figure 4. (a) CO₂ enrichment ratios (E/A) for basal area and basal area increment (BAI) of *Quercus ilex* trees in the vicinity of the Rapolano spring, Italy, and an adjacent control site. Basal area increments are presented as the 3-year running average. The regression line for BAI beginning at year 9 is: $E/A = -0.007 \times \text{age} + 1.36$; $R^2 = 0.09$. (b) Cumulative increase in basal area of CO₂-enriched trees compared to trees in ambient CO₂. Data courtesy of S. Hättenschwiler from experiment described in Hättenschwiler *et al.* (1997a).

annual growth per unit leaf area is a sustained response to CO₂ enrichment, albeit at somewhat less than the average value in Table 4. As a result of this sustained response, the cumulative gain in basal area (biomass) attributable to CO₂ enrichment increased with age and was not simply the result of the early stimulation of growth (Fig. 4b). Whether the unexplained decline in response in the last several years of the record at both Rapolano and Laiatico is the result of some aspect of stand development that will eventually lead to a complete loss of the CO₂ response, or a relatively short-term environmental fluctuation that will average out over time, cannot be determined. Hence, even with this much longer record of CO₂ response than has been available before, it remains difficult to predict the response in future decades. Nevertheless, these important

data sets from the CO₂ springs substantially extend the observation that the stimulation of tree growth by elevated CO₂ can be sustained over time under field conditions.

Allocation below ground

The allocation of carbon to below-ground tissues, and the growth, physiological activity and death of roots that results, are key points of intersection between the carbon cycle and the water and nutrient cycles. If experiments on tree responses to elevated CO₂ are to have relevance to forest ecosystem responses, there must be consideration given to the responses of root systems and associated below-ground processes. Unfortunately, of course, root responses are most difficult to study, and the inherent limitations in experimental approaches have meant that most of the observations are single observations at the end of an experiment, which is clearly problematic for such a dynamic system. The increasing use of minirhizotron systems has enabled more frequent observations, but the data can be difficult to quantify.

Earlier studies based on the responses of potted tree seedlings generally concluded that the ratio of root mass to shoot mass increases in elevated CO₂ (Oechel & Strain 1985), although perhaps only in low nutrient conditions (Eamus & Jarvis 1989; Bazzaz 1990). There are many problems with the measurement and interpretation of root-to-shoot ratio (Stulen & den Hertog 1993; Norby 1994), and past generalizations probably have little relevance to the issues of tree responses. It is especially important to separate the response of woody root mass from that of fine roots (Norby 1994). On the one hand, an increase in woody root mass implies storage of carbon just as an increase in bole wood does, but this cannot be surmised from young seedlings in which all of the roots are small. On the other hand, changes in whole-root system mass of older saplings or trees will tell us little about fine-root mass or turnover. Because of their much higher turnover rate, a large amount of carbon may be allocated to the production of fine roots, but the standing crop of fine roots can be a small percentage of the whole root mass. Nevertheless, the production and turnover of fine roots are critical processes linking plant response to soil response. Fine roots are the mechanism for nutrient uptake from the soil, the platform for microbial activity related to nutrient turnover, and the source of much of the carbon influx to soil (Norby 1994). Hence, we shall consider the experimental evidence for woody roots and fine roots separately.

Only a few multiyear studies of trees in elevated CO₂ have ended with a complete destructive harvest, so the data set on woody root response to elevated CO₂ is small. There was no significant effect on root-to-shoot ratio in *Liriodendron tulipifera* (Norby *et al.* 1992), *Quercus alba* (Norby *et al.* 1995), *Betula pendula* (Rey & Jarvis 1997), *Pinus taeda* (Tissue *et al.* 1997), *Pinus ponderosa* (Walker *et al.* 1997), or *Fraxinus excelsior*, *Quercus petraea* and *Pinus sylvestris* (Crookshanks *et al.* 1998). Static measures of root-to-shoot ratio may mask important treatment

effects on allocation that are confounded with developmental changes in allocation. Allometric analyses can be a more powerful method for examining allocation shifts. Tissue *et al.* (1997) found no effects of CO₂ on any allometric coefficients, including those describing root-shoot relations. Norby (1994) saw no effect of CO₂ on root-shoot allometry in *L. tulipifera*, but the allometric coefficient for *Q. alba* increased with increasing CO₂. Given the large root mass of many trees, such a shift could lead to underestimates of a CO₂ effect on total carbon storage based only on above-ground mass. For example, if the CPI for *B. pendula* is calculated to include the biomass increment for the stump and coarse root in addition to stem and branch production, the CO₂ effect on CPI increases from 9% (Table 4) to 21%.

In most field studies in which fine-root density (mass of roots per unit ground area) has been measured, fine roots have been shown to be especially responsive to CO₂. In the six studies represented in Fig. 5, fine-root density increased from 60 to 140% in elevated CO₂. Fine-root mass production also increased by 135% in 3-year-old *Pinus sylvestris* (Janssens *et al.* 1998), and fine root length density increased 63% in an oak-palmetto ecosystem (Day *et al.* 1996). Fine-root length production in *Fraxinus excelsior*, *Quercus petraea*, and *Pinus sylvestris* was increased by 95–240% in elevated CO₂ (Crookshanks *et al.* 1998). Although the direct impact of an increase in fine-root mass on whole-plant mass is small, it could nevertheless be important to longer-term ecosystem response. Increased fine-root density could, for example, support increased rates of nutrient uptake or stimulate increased rhizosphere activity. Although these static measures of fine root density tell us nothing about the total carbon flux to fine roots, there

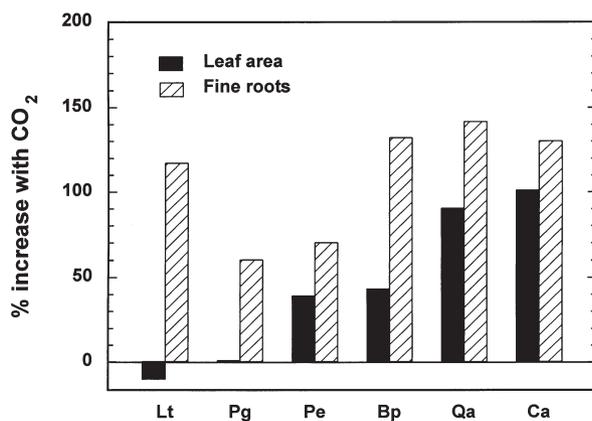


Figure 5. Relative effect of elevated CO₂ (percentage increase) on fine-root density and leaf area of trees exposed to elevated CO₂ in field experiments. Data are arranged in order of increasing effect of CO₂ on leaf area. Lt, *Liriodendron tulipifera* (Norby *et al.* 1992); Pg, *Populus grandidentata* (Zak *et al.* 1993); Pe, *Populus deltoides* × *P. nigra* (Pregitzer *et al.* 1995; Curtis *et al.* 1995); Bp, *Betula pendula* (Rey & Jarvis 1997); Qa, *Quercus alba* (Norby *et al.* 1995); Ca, *Citrus aurantium* (Idso & Kimball 1992b; Idso, Wall & Kimball 1993c).

is a presumption that increased fine root density indicates increased turnover as well, and root turnover is a mechanism for additional carbon to enter long-lived soil pools.

The large percentage increase in density of small roots (< 7 mm diameter) in *Liriodendron tulipifera* relative to the nonsignificant increase in whole-plant dry mass and decrease in leaf area (Norby *et al.* 1992) apparently confirmed the suggestion from a previous growth-chamber experiment (Norby & O'Neill 1991) that an important CO₂ response in field-grown trees could be a shift from leaf production to fine-root production. Such a mechanism could imply a shift in the tree's functional balance between carbon acquisition versus water and nutrient acquisition. In all of the studies represented in Fig. 5, the stimulation of fine-root density exceeded that of leaf area, and in all but *Citrus aurantium*, the relative response of fine roots also exceeded that of the whole plant. These observations suggest that stimulation of fine-root production may be a specific response to elevated CO₂, not simply a proportionate component of larger plants. Generally, the disparity between fine-root and leaf area response was smaller in those experiments in which leaf area showed the greatest response (the right end of the *x*-axis).

As discussed previously, the increase in LAI observed when open-grown trees are exposed to elevated CO₂ cannot be expected to persist indefinitely as a tree grows into a forest canopy. Likewise, the increase in fine-root density can be assumed to saturate as the soil volume becomes fully occupied. These static measures of fine-root density and leaf area do not predict whether a sustained increase in fine root to leaf area ratio is likely. It should, then, be important to look at the effect of CO₂ on fine roots in relation to the dynamics of the response of the rest of the plants. The use of minirhizotrons has allowed such analyses. Pregitzer *et al.* (1995) found that fine-root growth and mortality were more responsive to CO₂ than was leaf growth throughout their 1-year study, and data from a single destructive harvest would have been very misleading. Tingey *et al.* (1996) related fine-root dynamics of *Pinus ponderosa* to shoot growth dynamics over three growing seasons. Fine-root area density initially increased one to two-fold in elevated CO₂, but did not continue to increase as shoot growth continued. The ratio of fine roots to leaf area declined with time, and there was no effect of CO₂ on this ratio, although N fertilization did initially decrease the ratio.

Although there may well be differences between species or sites in the relative response of fine roots, the more rigorous observations afforded by periodic observations through minirhizotrons do not support the premise that there is a specific stimulation by elevated CO₂ of fine-root density or a shift in the functional balance between roots and foliage that is sustained over time. Nevertheless, it is important that fine-root production is enhanced at least to the same extent as that of the rest of the tree. A greater emphasis on fine-root turnover, instead of static measures of fine-root density, will help to reveal the potential importance of fine-root responses to whole-system function and carbon budget. Observations on the

horizontal (Thomas *et al.* 1996) and vertical distribution of fine roots and root carbon in soil through minirhizotron observation and quantification of mycorrhizal colonization (Rygiewicz *et al.* 1997; Runion *et al.* 1997) may make additional links to biogeochemical cycling.

NUTRIENT CYCLING

The importance of nutrient cycling as a control or modifier of CO₂ responses has been long recognized, and the focus has been mostly on nitrogen. Kramer (1981) questioned whether trees whose growth is limited by insufficient N in an unmanaged forest would respond to increased CO₂. Ecosystem models have strongly implicated N interactions as critical to the long-term response of forests to increasing CO₂. Models with strong links between the nutrient cycle and plant production generally predict smaller increases in production because of constraints imposed by N supply. Nitrogen limitation does not completely constrain the NPP response, however, because of internal recycling and seasonality in the limitation (McGuire *et al.* 1997). Various models differ in how N interactions are expressed, and comparison of several models indicated that these differences were the dominant factor in the prediction of the effect of CO₂ on net primary productivity (Ruimy *et al.* 1999). Despite many observations of N concentrations in CO₂-enriched trees and experimental manipulations of CO₂-N interactions, it is uncertain how N cycles will change with CO₂ enrichment and how those changes will influence the carbon cycle. The problem again is one of scale. To what extent can the nutrient budget of a tree seedling growing in a pot provide relevant data for the nutrient budget operating in a mature forest? The responses of trees with roots growing in and influencing unconstrained and unmanipulated soil, and with nutrients mobilized out of senescing leaves, stored in perennial tissue, and remobilized again in the next growing season, may come closer to the nutrient dynamics of a forest.

Foliar nitrogen concentration

The critical points of intersection between the carbon budget (as altered by elevated CO₂) and N cycling include the physiological demand of the tree for N and the annual rate of N uptake from the soil. Physiological demand can be thought of as the amount of N needed to sustain sufficient levels of enzymes for vital growth processes, such as the large requirement for N to maintain rubisco and other photosynthetic enzymes. Nitrogen shortages induced by accelerated growth in elevated CO₂ could cause lower concentrations of N in leaves, which would be expected to reduce the rate of photosynthesis (Field & Mooney 1986) but for the compensating effect of higher internal CO₂ concentration. Compilations of the data from many studies with potted seedlings have shown reductions in foliar [N] to be a common response to CO₂ enrichment (McGuire *et al.* 1995; Curtis 1996; Cotrufo *et al.* 1998). Hence, it is widely thought that enhanced photosynthesis and growth will not

be sustained because of N limitations, despite the substantial evidence and analyses to the contrary (Drake *et al.* 1997).

The summary of foliar [N] responses to elevated CO₂ in field-grown trees (Table 5) shows considerable variation, from an 20% increase in [N] to a 35% decrease, with an overall average decline of 11% in gymnosperms and 14% in angiosperms. These averages are less than the average values resulting from analyses of larger data sets that include potted tree seedlings (21%, McGuire *et al.* 1995; 16%, Curtis & Wang 1998; 16%, Cotrufo *et al.* 1998). There appears to be an effect of tree age (or duration of exposure) in that the average percentage reduction is lower in seedlings more than 2 years old, which explains the larger effect reported in previous data syntheses. However, the influence of plant age varies considerably between studies. In one study on *Pinus ponderosa*, there was a decline in the effect of CO₂ effect on foliar [N] with seedling age (e.g. Johnson, Ball & Walker 1997), but in other studies there was no consistent pattern (e.g. Ceulemans *et al.* 1996; Runion *et al.* 1997; Tissue *et al.* 1997). In the truly long-term studies in the natural CO₂ springs in Italy, the effect of elevated CO₂ was negative in one species and slightly positive in another (Körner & Miglietta 1994). Only two studies reported on the effects of soil N status on response to CO₂ (Pregitzer *et al.* 1995; Johnson *et al.* 1997), and again the results were inconsistent. In the *P. ponderosa* study, there were no consistent effects of N fertility on either foliar [N] itself or the response to elevated CO₂ (Johnson *et al.* 1997), whereas in the *Populus* study, both N fertility and CO₂ strongly affected foliar [N], the CO₂ effect being more pronounced at lower N fertility (Pregitzer *et al.* 1995). When all the data are plotted together (Fig. 6), the slope of the line of foliar [N] at elevated versus ambient CO₂ (0.89) is significantly less than 1, and the intercept (4.3 mg g⁻¹) is not significantly different from 0. Thus, this model would predict that the effect of elevated CO₂ is less (in absolute terms) at lower foliar [N]. Reductions in [N] can often be explained by a dilution effect of increased structural or nonstructural carbon in CO₂-enriched leaves (increased leaf mass per unit area) (Epron *et al.* 1996). Although N concentration on a leaf area basis (g N m⁻²) could not be determined for all of the studies in Table 5, the average decline was clearly much less than the decline in mass-based [N], especially after the first year of exposure (Table 5). In a meta-analysis of all experimental data on trees (Curtis 1996), there was no effect of CO₂ on N per unit leaf area, although mass-based [N] was reduced. This result supports the contention that the apparent decline in foliar N is more a function of the carbon economy of the leaf than a real decline in N.

Nitrogen uptake

Increased uptake of N from soil could allow N-deficient forests to respond to elevated CO₂ or could forestall impending N deficiency. Elevated CO₂ could facilitate increased uptake by stimulating root growth and soil

Table 5. Responses of foliar N concentration to elevated CO₂ in various open-top chamber studies. Entries grouped together are from a common experiment. When multiple measurements were reported, the data shown are averages across growing seasons. Nitrogen concentration on a leaf-area basis was calculated using reported values for leaf mass per unit area if necessary

Species	Group	Exposure duration	N mg g ⁻¹		% diff.	N mg g ⁻¹		% diff.	Reference
			Amb. CO ₂	Elev. CO ₂		Amb. CO ₂	Elev. CO ₂		
<i>Gymnosperms</i>									
<i>Picea abies</i>		2 years	24.8	23.9	-4				Le Thiec <i>et al.</i> 1995
<i>Picea abies</i>		16 months	15.8	14.0	-11				Marek <i>et al.</i> 1995
<i>Pinus palustris</i>		<1 year	8.9	6.9	-23				Runion <i>et al.</i> 1997
		1-2 years	10.7	8.2	-24				
<i>Pinus ponderosa</i>	Low N	1 year	17.2	13.0	-24				
	Low N	2 years	17.6	16.3	-7				
	Low N	3 years	12.1	10.7	-12				Johnson <i>et al.</i> 1997
	Medium N	1 year	17.4	11.7	-33				
	Medium N	2 years	17.1	15.2	-11				
	Medium N	3 years	12.0	12.4	3				
	High N	1 year	16.0	11.9	-26				
	High N	2 years	18.9	14.8	-22				
	High N	3 years	12.7	11.9	-6				
<i>Pinus radiata</i>	Current needles	1 year	11.4	17.3	20		1.46	1.96	Hogan <i>et al.</i> 1996
<i>Pinus radiata</i>	Current needles	4 years	15.3	15.0	-2		1.23	1.32	Turnbull <i>et al.</i> 1998
	1-year-old needles	4 years	11.2	9.8	-12		0.90	0.80	
<i>Pinus sylvestris</i>	Current needles	15 months	11.6	11.3	-3		3.20	2.90	Kellomäki & Wang 1997b
	1-year-old needles	15 months	11.0	10.6	-4		3.30	3.40	
<i>Pinus taeda</i>		<1 year	22.3	19.7	-12		1.08	1.08	Tissue <i>et al.</i> 1997
<i>Pinus taeda</i>		1-2 years	16.0	13.6	-15		0.81	0.88	Lewis <i>et al.</i> 1996
		2-4 years	16.9	14.8	-12				
<i>Angiosperms</i>									
<i>Acer rubrum</i>	Ambient temp.	15 months	25.6	19.1	-25		1.41	1.23	Norby, unpublished data
	Ambient temp.	25 months	21.5	20.4	-5		1.20	1.25	
	Ambient temp.	36 months	23.6	21.5	-9		0.89	0.77	
	Elevated temp.	15 months	20.7	20.0	-3		1.31	1.35	
	Elevated temp.	25 months	20.6	19.7	-4		1.20	1.21	
	Elevated temp.	36 months	20.1	19.3	-4		0.80	0.78	
	Ambient temp.	15 months	22.5	18.2	-19		1.25	1.23	
	Ambient temp.	26 months	20.5	17.8	-13		0.99	0.94	
	Ambient temp.	35 months	19.1	16.9	-12		0.69	0.72	
	Elevated temp.	15 months	21.2	20.3	-4		1.35	1.38	
	Elevated temp.	26 months	20.1	17.1	-15		1.12	0.98	
	Elevated temp.	35 months	18.8	16.4	-13		0.74	0.70	
<i>Acer rubrum</i>	High light	3 months	11.4	8.1	-29		0.61	0.55	Kubiske & Pregitzer 1996
	Low light	3 months	22.0	18.0	-18		0.57	0.48	
<i>Betula papyrifera</i>	High light	3 months	26.3	24.3	-8		1.99	1.45	
	Low light	3 months	29.3	24.7	-16		0.88	0.75	
<i>Quercus rubra</i>	High light	3 months	19.8	16.6	-16		2.11	1.31	
	Low light	3 months	26.1	21.2	-19		0.96	0.98	

Table 5. Continued.

Species	Group	Exposure duration	N mg g ⁻¹		% diff.	N mg g ⁻¹		% diff.	Reference
			Amb. CO ₂	Elev. CO ₂		Amb. CO ₂	Elev. CO ₂		
<i>Acer saccharum</i>		12 months	27.2	24.8	-9				Roth <i>et al.</i> 1998
<i>Populus tremuloides</i>		12 months	32.8	28.0	-15				
<i>Alnus glutinosa</i>		3 months	30.8	25.8	-16	1.76	1.94	10	Vogel & Curtis 1995
<i>Betula pendula</i>		3-4 years	25.1	23.4	-6	1.26	1.72	13	Rey & Jarvis 1998
<i>Citrus aurantium</i>		2-4 years	22.1	18.9	-15	2.94	2.77	-6	Peñuelas <i>et al.</i> 1997
		4-6 years	21.9	19.8	-10	2.39	2.23	-7	
		6-8 years	21.0	19.9	-5	2.54	2.45	-4	
<i>Lindera benzoin</i>		3 months	27.5	25.9	-6	0.73	0.73	0	Cipollini <i>et al.</i> 1993
<i>Liriodendron tulipifera</i>		2-3 years	20.7	13.7	-34	1.37	1.04	-24	Wüllschlegel <i>et al.</i> 1992a Norby <i>et al.</i> 1992
<i>Nothofagus fusca</i>		1 year	19.7	17.5	-11	2.37	2.24	-5	Hogan <i>et al.</i> 1996
<i>Populus deltoides</i>	Low N	< 1 year	23.0	17.0	-26				Curtis <i>et al.</i> 1995
× <i>P. nigra</i>	High N	< 1 year	33.9	33.2	-2				Pregitzer <i>et al.</i> 1995
<i>Populus deltoides</i>		1 year	17.9	12.3	-31	1.30	0.98	-25	Ceulemans <i>et al.</i> 1996
× <i>P. nigra</i>		2 years	18.6	15.3	-18				
<i>P. trichocarpa</i>		1 year	15.3	11.9	-22	0.97	0.92	-5	
× <i>P. deltoides</i>		2 years	18.4	11.9	-35				
<i>Quercus alba</i>		29 months	14.8	12.9	-13	1.08	1.13	5	Norby, unpublished data
		40 months	16.6	14.8	-11	1.21	1.30	7	
<i>Quercus alba</i>		37 months	23.0	19.0	-17				
<i>Quercus ilex</i>		> 100 years	12.2	13.2	8				Williams <i>et al.</i> 1998
<i>Quercus pubescens</i>		> 100 years	19.7	16.8	-15				Kömer & Miglietti 1994
<i>Quercus rubra</i>		2 years	28.5	27.4	-4				Le Thiec <i>et al.</i> 1995
Gymnosperms					-11.4 ± 11.8			-4.6 ± 15.3	
Angiosperms					-13.6 ± 9.3			-6.1 ± 11.6	
All species		< 1 year			-15.8 ± 8.0			-10.4 ± 15.3	
All species		1-2 years			-14.4 ± 12.3			-0.7 ± 14.8	
All species		> 2 years			-9.7 ± 7.9			-2.9 ± 9.3	

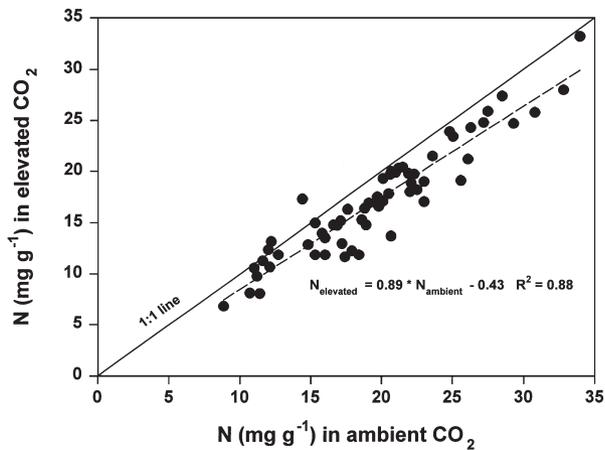


Figure 6. Nitrogen concentration (mg N g^{-1}) of leaves of trees grown in ambient CO_2 compared to leaves of trees grown in elevated CO_2 . All of the data are from trees (seedlings, saplings, and mature) rooted in the ground and exposed to CO_2 under field conditions, but the data encompass a wide range of species, interactive treatments, and exposure duration as shown in Table 5.

exploration, as shown in many seedling studies (Norby, O'Neill & Luxmoore 1986; Walker *et al.* 1995), or by increasing N availability through stimulation of N mineralization (e.g. Körner & Arnone 1992; Zak *et al.* 1993) or N_2 fixation. As discussed above, fine-root density has increased in field studies, and the increase generally exceeded that of leaf area, suggesting a potential improvement in the supply of N compared to demand, especially if root uptake capacity and mycorrhization are also stimulated. But the question remains as to whether fine root density, or the ratio of fine roots to leaf area, will continue to be enhanced after the soil is fully occupied by roots.

The data on N mineralization are equivocal. Zak *et al.* (1993) showed that elevated CO_2 caused increases in labile C and N in rhizosphere soil from *Populus grandidentata* seedlings. The authors posed a conceptual model whereby elevated CO_2 creates a positive feedback on soil C and N dynamics and tree growth because of increased carbohydrate allocation and, consequently, increased N availability in the rhizosphere. Curtis *et al.* (1994) report data from later studies of *P. grandidentata* and *P. deltoides* \times *P. nigra* supporting this model, at least under conditions with very low soil organic matter and N mineralization potential. On the other hand, the addition of labile organic C with low C/N ratio is known to immobilize available N (Paul & Clark 1989).

Few of the field studies in Table 1 have reported N uptake. The five data sets that we can compare (Fig. 7) are the 3-year study of *Pinus ponderosa* (Johnson *et al.* 1997), the 3-year study of *Liriodendron tulipifera* (Norby *et al.* 1996), the 4-year study of *P. taeda* (Tissue *et al.* 1997), the 1-year study of *Populus grandidentata* (Curtis *et al.* 1994), and the 4-year study of *Betula pendula* (Rey & Jarvis 1997, 1998). Larger trees with larger root systems can be assumed to take up more N. The important question is whether N

uptake increases commensurately with growth. This analysis is more difficult because a decline in whole-plant N concentration is an expected consequence of accelerated ontogeny, confounding any direct influence of CO_2 on [N] (Coleman, McConnaughay & Bazzaz 1993). Only in *B. pendula* did N uptake increase with CO_2 enrichment more than plant dry mass; hence, whole-plant [N] increased slightly in *B. pendula* and declined in the other three species. Nevertheless, N uptake increased substantially in all of the species except *Liriodendron tulipifera*. Was this increase attributable to (1) increased soil exploration, (2) increased mineralization, or (3) increased free-living N_2 fixation? In the *P. ponderosa* study, we know that N mineralization was initially reduced and then unaffected by elevated CO_2 . Increased soil exploration can be invoked in any of the three studies where N uptake was increased since all experienced an increase in root biomass. An important question arises, however, as to whether mature, closed-

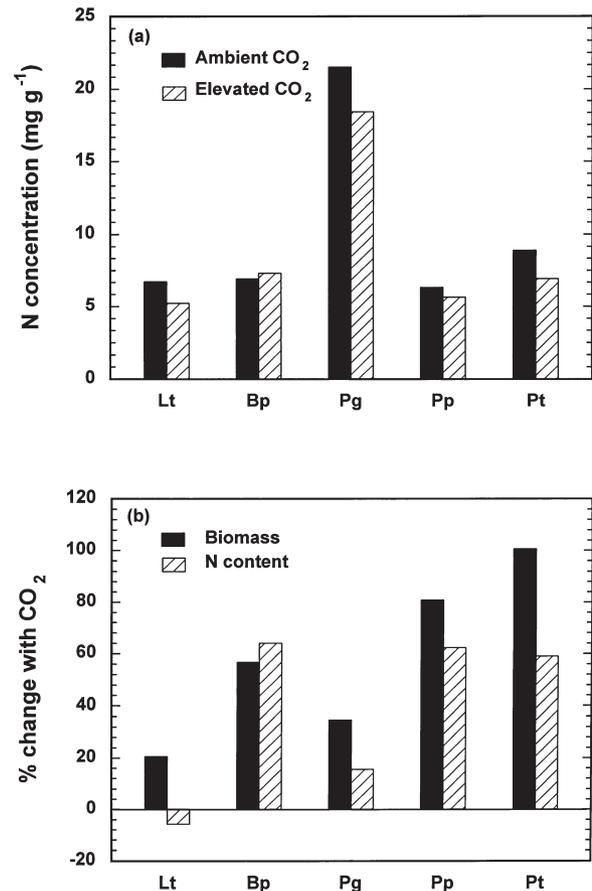


Figure 7. (a) Nitrogen concentration (mg N g^{-1}) of the whole plant (leaves, stems, and roots) of five tree species grown in ambient or elevated CO_2 , and (b) the relative effect of CO_2 enrichment (percentage change from ambient) of whole-tree N content and dry mass. Lt, *Liriodendron tulipifera* (Norby *et al.* 1996); Bp, *Betula pendula* (Rey & Jarvis 1997, 1998); Pg, *Populus grandidentata* (Curtis *et al.* 1994 and personal communication); Pp, *Pinus ponderosa* (Johnson *et al.* 1997); Pt (*Pinus taeda*, Tissue *et al.* 1997).

canopy forests, where root systems have been exploring the soil for decades, can increase N uptake by increasing root biomass and soil exploration. The answer to this question is of vital importance in assessing the potential for landscape-scale forest response to elevated CO₂, as most forest ecosystems are at a closed-canopy stage.

None of these CO₂ studies has measured free-living N₂ fixation, but studies by Bormann *et al.* (1993) suggest that this can be a major source of N in *Pinus* species. The effect of elevated CO₂ on this free-living N₂ fixation constitutes a 'free lunch' for those species in which it might occur, especially with the increased below-ground inputs of carbohydrates that are often accompanied by elevated CO₂ (Zak *et al.* 1993). Similarly, enhancement by elevated CO₂ of N₂ fixation by certain lichens could add a small amount of additional N to some forest ecosystems (Norby & Sigal 1989).

Carbon–nitrogen linkages

Modelling studies have suggested that over a time scale of decades there will be significant negative feedback on tree growth because of a decline in decomposition and N cycling rates related to lower-quality litter from CO₂-enriched trees. (Strain 1985; Rastetter *et al.* 1992). A decline in N cycling could be a significant factor in mature forests where > 80% of N taken up by trees every year is recycled (Cole & Rapp 1981). Slow decomposition and forest floor build-up have been connected to progressive N deficiency with stand age in *Pseudotsuga menziesii* ecosystems (Turner 1977). For logistic reasons, there have been no studies of the effects of elevated CO₂ on N cycling in forests. Several researchers have approached the problem, however, by investigating effects on litter quality or decomposition rate, often in the laboratory. The results of these studies have been mixed and generally inconclusive. Cotrufo, Ineson & Rowland (1994) found significant effects of elevated CO₂ on litter quality, decomposition, and N availability in senescent *Fraxinus excelsior*, *Betula pubescens*, and *Acer pseudoplatanus* leaves in a laboratory study. Randlett *et al.* (1996) found no effect of elevated CO₂ on decomposition or N mineralization of leaves of *Populus deltoides* × *P. nigra*. O'Neill & Norby (1996) reported no effect of CO₂ enrichment on litter quality or decomposition (mass loss) of *Liriodendron tulipifera* leaves. Reviewing the studies published at that time, O'Neill & Norby (1996) concluded that most of the reported cases of a CO₂ effect on litter quality (i.e. on the C:N or lignin:N ratio) occurred in potted seedlings in which the litter quality was substantially different from that of trees in the field, suggesting a possible artifact of the nutrient regimen in the pot. Elevated CO₂ has not been shown consistently to reduce leaf-litter quality of field-grown trees. This conclusion has been supported by observations of oak leaf litter in the vicinity of CO₂ springs in Italy. There were no statistically significant differences in N concentration, C:N ratio, or mass loss of senescent *Quercus pubescens* leaves from a high

CO₂ area compared to a reference area (Ineson & Cotrufo 1997), although some trends were noted and discussed. More extensive observations of *Q. pubescens* and *Q. cerris* leaf litter at a different CO₂ spring led Gahrooee (1998) to the conclusion that elevated CO₂ has no impact on litter chemistry of Mediterranean *Quercus* species, and consequently litter turnover is not affected. Nevertheless, the importance of this linkage between the carbon cycle and the N cycle as a regulator of long-term forest productivity makes it mandatory to consider possible effects of CO₂ in longer-term experiments.

Herbivorous insects are an important contributor to the fluxes of carbon and nitrogen in forest ecosystems. The observations of lower [N] in leaves of plants grown in elevated CO₂ led to the suggestion that the behaviour of herbivores feeding on those leaves might be affected (Lincoln, Fajer & Johnson 1993). Experiments in open-top chambers made possible more extensive field trials of herbivore interactions under field conditions. Pine sawfly (*Neodiprion lecontei*) larvae consumed more needle tissue from *Pinus taeda* trees in elevated CO₂ to compensate for the lower [N] of the foliage compared to that of ambient-grown trees (Williams, Lincoln & Thomas 1997). Larvae of the gypsy moth (*Lymantria dispar*) had reduced growth, prolonged development, and increased consumption when feeding on leaves of CO₂-enriched *Populus tremuloides* (Lindroth *et al.* 1997), related to marginally reduced [N] and increased content of condensed tannins relative to foliage in ambient CO₂. However, there was no significant effect of foliage quality on final pupal mass or female fecundity. The content of condensed tannins also increased in *Pinus palustris* grown in elevated CO₂ (Pritchard *et al.* 1997). The growth rate of early instar larvae of gypsy moth was significantly reduced when they were presented with young, expanding *Quercus alba* leaves from trees in elevated CO₂ (Williams, Lincoln & Norby 1998). The leaves had lower leaf N content but similar nonstructural carbohydrate and phenolic content compared to leaves from trees grown in ambient CO₂. The growth rate of forest tent caterpillar (*Malacosoma disstria*) larvae was not affected, nor were the consumption rates or growth rates of later instars of either insects that were fed older leaves (Williams *et al.* 1998). In a different study, however, the same species exhibited reduced growth and food processing efficiencies when fed foliage from CO₂-enriched *Acer saccharum* and *Populus tremuloides* trees relative to insects fed ambient-grown foliage (Roth *et al.* 1998). These various observations suggest the possibility that the interactions between trees and herbivorous insects could change as the atmospheric CO₂ concentration increases, but that the responses cannot be predicted simply from the effects of CO₂ on foliar [N]. Although CO₂ effects on herbivory could have important ramifications on forest health, forest productivity and nutrient cycling, there is not yet any framework for integrating these experimental observations with the population dynamics of the insect, as would be necessary for an assessment of the impact on ecosystem productivity.

WATER

Stomatal conductance in response to elevated CO₂

The short-term exposure of plants to elevated CO₂ has long been known to decrease stomatal conductance in a range of herbaceous crops and woody species. The reported magnitude of this response varies from a 40% reduction in stomatal conductance for 16 C₃ and nine C₄ crops (Morison 1985) to a 27% reduction for 20 species of woody plants grown in pots and exposed to atmospheric CO₂ enrichment (Field, Jackson & Mooney 1995). At issue, however, is whether general reductions in stomatal conductance can be expected for both broadleaved hardwoods and conifers exposed to elevated CO₂ in long-term studies conducted under field conditions. Surprisingly, recent studies indicate little or no effect of atmospheric CO₂ on stomatal conductance. There was, for example, no effect of a doubling of CO₂ on stomatal conductance in two hybrid *Populus* clones (Will & Ceulemans 1997), no effect in *Quercus alba* and *Liriodendron tulipifera* seedlings (Wullschleger *et al.* 1992b), only modest reductions (up to 15%) in *Quercus alba* and *Liriodendron tulipifera* saplings (Gunderson *et al.* 1993), a 14% reduction in *Pinus sylvestris* (Wang & Kellomäki 1997), a small to no significant effect in *P. taeda* (Ellsworth *et al.* 1995; Teskey 1995; Tissue *et al.* 1997), and only slight (10%) reductions in *Picea abies* (Dixon *et al.* 1995).

These field studies indicate that the sensitivity of stomatal conductance to elevated CO₂ is far less than that reported for a range of herbaceous species and trees in earlier growth-chamber studies. Saxe, Ellsworth & Heath (1998) suggest that the magnitude of stomatal response to elevated CO₂ is indeed smaller in trees than in crops and herbs, and that differences also exist between woody deciduous and coniferous species. According to their analysis, most conifers show a small or non-significant reduction in stomatal conductance upon exposure to elevated CO₂ in the field, while stomata of deciduous trees show a stronger response. Herbaceous crops and grasses by comparison almost always show a much larger CO₂-induced reduction in stomatal conductance than do trees. While the mechanisms that mediate this differential response of stomata among herbaceous crops, grasses, and deciduous and coniferous trees are not well understood, data collected from recent field studies emphasize that assumptions concerning the perceived sensitivity of stomatal conductance to atmospheric CO₂ enrichment must be re-evaluated. These revised assumptions will not only influence model simulations of whole-plant transpiration and stand water use (Martin 1992), but also help to refine model estimates of evapotranspiration and thereby improve our ability to predict the role of CO₂-induced biotic feedbacks in modifying regional and global climate (Henderson-Sellers, McGuffie & Gross 1995; Pollard & Thompson 1995; Sellers *et al.* 1996).

Transpiration and canopy water use

Much of our interest in the response of stomatal conductance to atmospheric CO₂ enrichment relates to a need for quantitative estimates of leaf transpiration. Few studies report rates of leaf transpiration, although one might conclude from the effects of elevated CO₂ on stomatal conductance that the response is likely to be small. Such was found by Teskey (1995) who, in addition to observing no effect of elevated CO₂ on stomatal conductance in his branch-bag studies of *Pinus taeda*, observed no effect of CO₂ on needle transpiration. Even when reductions in stomatal conductance are observed, there are reasons why these effects may not necessarily contribute to reductions in leaf-level transpiration. For example, CO₂-induced reductions in stomatal conductance and (at least temporally) transpiration should contribute to an increase in leaf temperature. This increase in leaf or needle temperature exerts negative feedback on transpiration, and rates of transpiration may therefore increase after partial stomatal closure. The complex interactions between stomatal conductance, transpiration and leaf temperature have been examined in agricultural studies (Idso *et al.* 1993b), but they have not been addressed experimentally for trees grown at elevated CO₂ concentration. This represents a major shortcoming of previous field experiments and such a deficiency should be remedied in future studies.

Field studies that document effects of elevated CO₂ on stomatal conductance and transpiration will be challenged to apply this knowledge at the scale of whole trees and canopies. This shift in focus from leaf-level determinants of transpiration to those operating at the scale of forest canopies will require that other non-stomatal processes be considered in the control of whole-tree water loss. Boundary layers that surround individual leaves and canopies are of critical importance and, especially for broadleaved species, will probably cause the reductions in the canopy transpiration caused by CO₂-induced stomatal closure to be smaller than would otherwise be inferred from single-leaf measurements. Studies to examine the response of large trees to elevated CO₂ and the implications of CO₂-induced alterations to leaf physiology and canopy biophysics are clearly needed. In this regard, free-air CO₂ enrichment (FACE) facilities and natural CO₂ springs offer unique opportunities to explore trade-offs between stomatal and boundary layer conductances in the control of whole-plant water use. Ellsworth *et al.* (1995) addressed these topics by quantifying canopy water use for *Pinus taeda* exposed briefly (8 d) to atmospheric CO₂ enrichment, as did Tognetti *et al.* (1996) for *Quercus pubescens* and *Q. ilex* at a CO₂ spring in central Italy. This latter study coupled leaf-level measurements of stomatal conductance, transpiration and leaf water potential with whole-tree estimates of sap velocity to compare water relations for trees growing in or near a natural CO₂ spring. Studies that use such a combination of leaf and whole-tree measurements should be expanded, and similar activities at existing FACE facilities and natural CO₂ springs should be encouraged.

Leaf and canopy controls of whole-tree water use will ultimately have to integrate a wide variety of CO₂-induced effects on plant growth, fine-root density and distribution and leaf area production. Enhanced root proliferation for trees grown at elevated CO₂ (Thomas *et al.* 1996; Tingey *et al.* 1997) or a preferential distribution of roots to deeper soil profiles (Day *et al.* 1996) may provide increased access to soil water. While this is an attractive hypothesis, it is doubtful that cause-and-effect relationships can easily be established. Larger plants with greater leaf area, or stands with greater LAI, are expected to offset or compensate for reductions in stomatal conductance and thereby contribute to higher rates of whole-plant water use. As previously discussed, the ability to increase leaf area per plant has been demonstrated in a number of field studies, although the response of LAI in a closed-canopy forest is unknown. The possibility of lower LAI in elevated CO₂ (Hättenschwiler & Körner 1998) can be interpreted as a morphological adjustment or mechanism of down-regulation that operates at the canopy scale, and therefore may have implications for tree water consumption.

WHERE DO WE STAND? WHERE ARE WE HEADING?

Experiments with trees will always be difficult. Trees live for a long time, grow to a large size, and exist in a complex environment of competing species and spatially and temporally variable resources. While it is clearly important to recognize the many problems in interpreting the data from small, young trees in a simplified environment (Lee & Jarvis 1995), and new larger-scale experiments will always be called for, it is also important that we search for innovative and perceptive ways of viewing the available data sets. We maintain that experiments completed with young trees in open-top chambers offer a rich source of information to guide the development of new experimental and modelling approaches.

How good were seedling studies?

A primary rationale for conducting CO₂ enrichment studies in open-top chambers was the need to determine if the responses observed in short-term studies with seedlings in greenhouses and growth chambers are sustained over several growing seasons under field conditions. This was a particularly important question with regard to trees, because so much of what describes tree growth relates to its perenniality — the storage and remobilization of carbon and nutrients from one year to the next, the exposure to many uncontrolled and constantly fluctuating environmental resources and stresses, and the large size resulting from cumulative growth over many years. The database of responses of trees to elevated CO₂ under field conditions is sufficient for us to assess, looking retrospectively, the value and robustness of conclusions from the earlier studies and, looking prospectively, the remaining questions and uncertainties that must be addressed in still larger-scale experiments.

Most of what was learned in seedling studies was qualitatively correct: photosynthesis is enhanced, N concentrations are reduced, plants are bigger at the end of the experiment. Quantitative comparisons are problematic because the range of response can be so large. Nevertheless, it seems safe to conclude that photosynthetic enhancement of tree leaves in the field is similar to (or greater than) that observed in seedling studies. Suggestions that photosynthetic enhancement would not be sustained — an important reason for conducting longer-term studies — turn out not to be valid in most cases. Down-regulation of leaf-level photosynthesis is not consistently observed in the field. Foliar [N] is reduced, at least on a leaf mass basis, but the reduction is less than was indicated in seedling studies, where artifacts of unbalanced nutrition were more likely to occur.

Attempts to compare growth responses are especially problematic, but reveal what are the important considerations for scaling. The average response of final dry mass (which is not the same as growth) of the field-grown trees is a 64% increase (log-adjusted) in elevated CO₂, which exceeds most compilations of the average response of all tree species (dominated by seedling studies). The larger apparent response in the field experiments may be a consequence of exponential growth operating over a longer period, magnifying any effect of CO₂ on growth rate. Our main objective should be to determine the effect of elevated CO₂ not on final dry mass but on growth rate — the parameter closer to annual increase in carbon storage. In the short-term studies that begin with seeds or small seedlings, differences in final dry mass should be indicative of differences in growth rates. In multiyear studies, however, growth rate can change considerably through time and in relation to plant development. An average response to CO₂ enrichment of this dynamic process, as represented by the difference in dry mass at a particular point in time, is not meaningful. Normalization of dry mass increases to a constant leaf area (the CPI) is one way to produce meaningful growth-rate data. The result, a 27% increase in CPI in elevated CO₂, is remarkably close to the most recent values for average growth increases in seedling studies (Wullschlegel *et al.* 1997a; Curtis & Wang 1998). Surprisingly, the seedling response may be a better predictor of long-term tree growth response than the simple averages from the field data.

Other predictions from the seedlings studies are less robust. Stomatal conductance was almost universally thought to be reduced by elevated CO₂ (although there were exceptions), but the responses of field-grown trees are less consistent and apparently less important. Leaf-litter quality is not altered by elevated CO₂ in the field as was suggested from controlled environment studies, perhaps because leaf senescence occurs under more natural conditions in the field. Increases in root-to-shoot mass ratio were widely predicted from seedling studies, but there is little indication that allocation is affected by CO₂ in the larger, older trees used in the field experiments. However, there appears to be a specific effect of CO₂ on fine-root mass, as was predicted from a few seedling studies.

The field studies summarized here have provided no reason to challenge accepted views on tree responses to elevated CO₂. Where there are discrepancies with previous understanding, the explanation does not lie in a fundamental difference in biology, but in experimental artifacts created by artificial nutrient regimens (e.g. confined roots, fixed N capital) or in the interactions of response with normal, predictable patterns of plant development. Both of these issues were, or should have been, recognized when the seedling studies were conducted, but the implications were sometimes ignored. It also appears that the research community was too ready to accept as dogma some of the trends observed in response to elevated CO₂ (e.g. litter quality is reduced, stomatal conductance is lower). Many exceptions to these trends have been observed in controlled-environment experiments, and the lack of consistency is now more apparent, but there is no evidence that the basic biology is different.

Can we predict forest responses?

The general concurrence between seedling studies and field studies, as well as the understanding of why there are discrepancies, improves the prospects for success in predicting the responses of larger trees in forests over much longer periods. There are, to be sure, many differences between the young trees in open-top chambers and forest trees (Lee & Jarvis 1995), and it is important to recognize the limitations of the current data set. These limitations are in three major areas: the over-riding influence of tree and stand developmental patterns, the lack of an ecosystem perspective in many of the measured responses, and scaling issues.

Interpreting the responses of trees in open-top chambers or in any other system without regard to developmental patterns will inevitably lead to false conclusions. Do trees use more water in elevated CO₂ even if stomatal conductance is reduced? They do if faster growth has produced a larger canopy, but this conclusion cannot be applied to a forest stand that has reached its maximum LAI. Do trees in elevated CO₂ take up more nutrients? Most of the trees in the open-top experiments had greater nutrient contents (but not concentrations) in elevated CO₂ because their root systems were larger, but this response is not relevant to a tree in a mature stand that has fully occupied the soil. These examples are not meant to suggest that we have learned nothing useful about water use or nutrient uptake but rather to emphasize the importance of separating functional responses from structural differences that are derived from developmental differences. Interpreting a growth response to CO₂ enrichment is a more difficult challenge because growth and development are so closely linked. A common conclusion following assessment of the likelihood of a sustained, long-term stimulation is that growth enhancement will decline with time, and the only lasting benefit of elevated CO₂ is the relatively small effect deriving from faster initial growth (Jarvis 1998). The long-term effect, it is thought, will be much less than that predicted from short-term experiments. This statement is difficult to evaluate

without defining what is the expected or baseline response. If the 'expected' outcome is a doubling of plant mass, as occurred in several experiments, then the response will almost certainly decline with time because those large increases are dependent on the compound interest of an increasing leaf area. If, however, the expected outcome takes account of developmental trends and assumes that the long-term CO₂ effect is the residual (by normalizing to constant leaf area), then there is no clear indication from the experimental data that the annual growth enhancement will decline from a value of about 25–30%.

Modelling exercises have indicated that ecosystem responses to elevated CO₂ will decline with time because of ecosystem-level feedbacks, particularly through the N cycle. An important limitation of the existing database of CO₂ experiments is that the potentially important feedback mechanisms cannot be fully evaluated for forest systems. The simple reason is that forest ecosystems are not the unit of study in open-top chamber experiments. Components of forest systems — individual trees, specific soil processes — are studied, and those studies provide useful input to ecosystem models, but the integration of those components requires a larger-scale experiment. Two examples of this limitation are the lack of a true nutrient cycle in the experimental systems and the absence of competing species in most experiments.

The failure to deal with specific scaling issues is another inevitable limitation of these experimental studies. Most experiments used only two concentrations of CO₂, and those that used additional levels did not have enough statistical power to resolve departures from linearity. It is highly likely, however, that most responses to CO₂ are non-linear (Körner 1995). Hence, our response data are only semi-quantitative, and in this review we usually referred only to 'elevated' CO₂ rather than to a specific concentration. The other important scaling issue is that some important controls on large-scale system response do not pertain to the smaller scale of the field experiments described here. A prominent example is the canopy boundary layer that strongly influences forest stand transpiration but is not so important in controlling plant transpiration in open-top chambers.

Can the current data set guide new experiments?

These limitations are not listed to cast doubt on the value of our existing data set. Instead, this analysis should provide a basis for new experiments that are being conducted at a larger scale. Free-air CO₂ enrichment (FACE) studies can move beyond many of the limitations of open-top chamber experiments: the basic unit of response can be a stand or ecosystem rather than an individual plant, the components of the plant–soil nutrient cycle are fully integrated, there can be a fully developed forest canopy, and different species can compete for resources. The forest stands within FACE arrays, however, will not replicate the forest of 50–100 years in the future — the plant material, soil development and land-use history will all be different, and a few

small plots of forest cannot be truly representative of an entire region or forest type. Instead, it is appropriate to think of the FACE experiments as experimental systems for testing specific, well defined hypotheses that will continue to guide the development of ecosystem models of long-term forest response. Those hypotheses should be developed based on the best understanding currently available on tree responses to elevated CO₂. Important hypotheses might include: (1) maximum LAI will increase in elevated CO₂ because shaded leaves deep in the canopy will be retained longer; (2) annual tree growth per unit LAI will continue to be enhanced by CO₂ after canopy closure; (3) fine-root density will not change in elevated CO₂ but fine-root turnover will increase; (4) down-regulation of tree growth responses will occur through long-term changes in the N cycle; (5) tree water use will be decoupled from any persistent CO₂ effects on stomatal conductance; (6) differential effects of CO₂ on competing species during establishment phase will alter long-term stand composition and productivity. There are, of course, many other possible hypotheses that are based on the current data and will increase the scale at which we understand forest response.

Not all important questions about forest response are amenable to FACE experiments, and other approaches need to be pursued simultaneously. The value of investigations in forests surrounding natural CO₂ springs has already been demonstrated (Hättenschwiler *et al.* 1997a), and despite their drawbacks (especially the problem of identifying an appropriate control site), the spring sites offer a unique opportunity to explore the long-term implications of the responses observed in shorter-term studies. Constructed microcosms offer the opportunity to manipulate species interactions and competition (Körner 1995), as long as artifacts through the below-ground environment are avoided. Environmental interactions that cannot currently be manipulated at the scale of a FACE experiment (e.g. air temperature) can still be explored in open-top chambers (Norby *et al.* 1997), although all of the scale-dependent provisos discussed in this review must be recognized. The interaction between temperature and CO₂ is an important parameter of global change, and therefore particularly relevant to explore. Many studies have shown extreme sensitivity of growth processes to rather small changes in growth temperature above or below the current ambient range (Ceulemans 1997). Open-top chamber experiments will continue to be useful in certain low-stature forest systems such as the oak-palmetto system in Florida (Day *et al.* 1996) and the natural Mediterranean macchia (Scarascia-Mugnozza *et al.* 1996), although such systems may not be very representative of more productive forests. This approach might represent a reasonable basis on which to extrapolate results obtained on a canopy of young trees (in a microcosm study, for example) to canopies of larger trees.

The influence of competition on forest stand development is well known, yet barely addressed in CO₂ research except at the scale of small constructed systems in containers. Can the knowledge we have gained at the tree level can be applied to the stand level, given the importance of

competition in real forests? In the presence of competing species, the response of the individual may be highly modified and not predict the response of communities (Bazzaz 1990). Every experiment with multiple species has shown differences in response to CO₂ (Körner 1995), and it is quite likely that the response of ecosystem productivity to rising CO₂ will result primarily from changes in species composition brought on by differential species responses to CO₂ (Bazzaz 1990). Nevertheless, there is not yet any basis for summarizing differential responses of tree species to CO₂ or for predicting the effect of elevated CO₂ on the outcome of competition in a regenerating forest. The role of competition is especially important with regard to leaf area and canopy development, which we have emphasized to be a critical uncertainty that hinders our ability to extrapolate from the current data set. Microcosms containing mini-stands of trees that reach a closed canopy status at an early stage might provide a feasible way to address some of these questions (Overdieck 1993).

As the research community moves on to a new generation of experiments, several things seem to be clear. There will be closer integration between experimental studies and ecosystem model development. The new experiments will advance our understanding of forest responses at a larger and more realistic scale than has previously been possible. We will inevitably learn that some of our conclusions from the current data set are wrong, while other conclusions will be supported. We may not be able to provide definitive answers about the global forest in a constantly changing atmosphere, but if the experiments are done correctly and the results are analysed with sensitivity to the inherent regulators and constraints on forest productivity, we shall continue to deepen our understanding while refining the questions.

ACKNOWLEDGMENTS

We thank Ana Rey, Y.-P. Wang, David Olzyck, Jud Isebrands and David Tissue, who shared with us their unpublished manuscripts. Special thanks to Y.-P. Wang who kindly provided unpublished annual estimates of gross photosynthesis and modelled estimates of growth and maintenance respiration for leaves, stems and roots of young birch trees, to Stephan Hättenschwiler for sharing the data used in Fig. 4, and to Peter Curtis for nitrogen data. Research was sponsored by the Global Change Research Program of the Environmental Sciences Division, U.S. Department of Energy, under contract number DE-AC05-96OR22464 with Lockheed Martin Energy Research Corp. This work is part of the Global Change and Terrestrial Ecosystems Core Project of the International Geosphere-Biosphere Programme. Publication no. 4859.

REFERENCES

- Amthor J.S. (1995) Terrestrial higher-plant response to increasing atmospheric [CO₂] in relation to the global carbon cycle. *Global Change Biology* **1**, 243–274.

- Arnore J.A. & Körner Ch. (1993) Influence of elevated CO₂ on canopy development and red–far-red ratios in 2-storied stands of *Ricinus communis*. *Oecologia* **94**, 510–515.
- Badeck F.W., Dufrêne E., Epron D., Le Dantec V., Liozon R., Mousseau M., Pontailler J.Y. & Saugier B. (1997) Sweet chestnut and beech saplings under elevated CO₂. In *Impacts of Global Change on Tree Physiology and Forest Ecosystems* (eds G.M.J. Mohren, K. Kramer & S. Sabate), pp. 15–25. Kluwer Academic Publishers, Dordrecht.
- BassiriRad H., Thomas R.B., Reynolds J.F. & Strain B.R. (1996) Differential responses of root uptake kinetics of NH₄⁺ and NO₃⁻ to enriched atmospheric CO₂ concentration in field-grown loblolly pine. *Plant, Cell and Environment* **19**, 367–371.
- Bazzaz F.A. (1990) Response of natural ecosystems to the rising global CO₂ levels. *Annual Review of Ecology and Systematics* **21**, 167–196.
- Bazzaz F.A., Miao S.L. & Wayne P.M. (1993) CO₂-induced growth enhancements of cooccurring tree species decline at different rates. *Oecologia* **96**, 478–482.
- Bormann B.T., Bormann F.H., Bowden W.B., Pierce R.S., Hamburg S.P., Wang D., Snyder M.C., Li C.Y. & Ingersoll R.C. (1993) Rapid N₂ fixation in pines, alder, and locust — evidence from the sandbox ecosystem study. *Ecology* **74**, 583–598.
- Carey E.V., DeLucia E.H. & Ball J.T. (1996) Stem maintenance and construction respiration in *Pinus ponderosa* grown in different concentrations of atmospheric CO₂. *Tree Physiology* **16**, 125–130.
- Ceulemans R. (1997) Direct impacts of CO₂ and temperature on physiological responses in trees. In *Impacts of Global Change on Tree Physiology and Forest Ecosystems* (eds G.M.J. Mohren, K. Kramer & S. Sabate), pp. 3–14. Kluwer Academic Publishers, Dordrecht.
- Ceulemans R., Jiang X.N. & Shao B.Y. (1995) Effects of elevated atmospheric CO₂ on growth, biomass production and nitrogen allocation of two *Populus* clones. *Journal of Biogeography* **22**, 261–268.
- Ceulemans R. & Mousseau M. (1994) Tansley Review no. 71: Effects of elevated atmospheric CO₂ on woody plants. *New Phytologist* **127**, 425–446.
- Ceulemans R., Shao B.Y., Jiang X.N. & Kalina J. (1996) First- and second-year aboveground growth and productivity of two *Populus* hybrids grown at ambient and elevated CO₂. *Tree Physiology* **16**, 61–68.
- Ceulemans R., Taylor G., Bosac C., Wilkins D. & Besford R.T. (1997) Photosynthetic acclimation to elevated CO₂ in poplar grown in glasshouse cabinets or in open top chambers depends on duration of exposure. *Journal of Experimental Botany* **48**, 1681–1689.
- Chaves M.M. & Pereira J.S. (1992) Water stress, CO₂ and climate change. *Journal of Experimental Botany* **43**, 1131–1139.
- Cipollini M.L., Drake B.G. & Whigham D. (1993) Effects of elevated CO₂ on growth and carbon/nutrient balance in the deciduous woody shrub *Lindera benzoin* (L.) Blume (Lauraceae). *Oecologia* **96**, 339–346.
- Cole D.W. & Rapp M. (1981) Elemental cycling in forest ecosystems. In *Dynamic Properties of Forest Ecosystems*. (ed. D. E. Reichle), pp. 341–409. Cambridge University Press, London.
- Coleman J.S., McConnaughay K.D.M. & Bazzaz F.A. (1993) Elevated CO₂ and plant nitrogen-use: is reduced tissue nitrogen concentration size-dependent? *Oecologia* **93**, 195–200.
- Cotrufo M.F., Ineson P. & Rowland A.P. (1994) Decomposition of tree leaf litters grown under elevated CO₂: effect of litter quality. *Plant and Soil* **163**, 121–130.
- Cotrufo M.F., Ineson P. & Scott A. (1998) Elevated CO₂ reduces the nitrogen concentration of plant tissues. *Global Change Biology* **4**, 43–54.
- Crookshanks M., Taylor G. & Broadmeadow M. (1998) Elevated CO₂ and tree root growth: contrasting responses in *Fraxinus excelsior*, *Quercus petraea* and *Pinus sylvestris*. *New Phytologist* **138**, 241–250.
- Cure J.D. & Acock B. (1986) Crop responses to carbon dioxide doubling: a literature survey. *Agricultural and Forest Meteorology* **38**, 127–145.
- Curtis P.S. (1996) A meta-analysis of leaf gas exchange and nitrogen in trees grown under elevated carbon dioxide. *Plant, Cell and Environment* **19**, 127–137.
- Curtis P.S., Vogel C.S., Pregitzer K.S., Zak D.R. & Teeri J.A. (1995) Interacting effects of soil fertility and atmospheric CO₂ on leaf growth and carbon gain physiology in *Populus × euramericana* (Dode) Guinier. *New Phytologist* **129**, 253–263.
- Curtis P.S. & Wang X. (1998) A meta-analysis of elevated CO₂ effects on woody plant mass, form, and physiology. *Oecologia* **113**, 299–313.
- Curtis P.S., Zak D.R., Pregitzer K.S. & Teeri J.A. (1994) Above- and belowground response of *Populus grandidentata* to elevated atmospheric CO₂ and soil N availability. *Plant and Soil* **165**, 45–51.
- Day F.P., Weber E.P., Hinkle C.R. & Drake B.G. (1996) Effects of elevated atmospheric CO₂ on fine root length and distribution in an oak–palmetto scrub ecosystem in central Florida. *Global Change Biology* **2**, 143–148.
- Dixon M., Le Thiec D. & Garrec J.P. (1995) The growth and gas exchange response of soil-planted Norway spruce [*Picea abies* (L.) Karst.] and red oak (*Quercus rubra* L.) exposed to elevated CO₂ and to naturally occurring drought. *New Phytologist* **129**, 265–273.
- Drake B.G., Azcon-Bieto J., Berry J., Bunce J., Dijkstra P., Farrar J., Gifford R.M., Gonzalez-Meler M.A., Koch G., Lambers H., Siedow J. & Wullschlegel S. (1999) Does elevated CO₂ concentration inhibit mitochondrial respiration in green plants? *Plant, Cell and Environment* **22**, 649–657.
- Drake B.G., Gonzalez-Meler M.A. & Long S.P. (1997) More efficient plants: a consequence of rising atmospheric CO₂? *Annual Review of Plant Physiology and Plant Molecular Biology* **48**, 609–639.
- Dvorak V. & Oplustilova M. (1997) Respiration of woody tissues of Norway spruce in elevated CO₂ concentration. In *Impacts of Global Change on Tree Physiology and Forest Ecosystems* (eds G.M.J. Mohren, K. Kramer & S. Sabate), pp. 47–51. Kluwer Academic Publishers, Dordrecht.
- Eamus D., Duff G.A. & Berryman C.A. (1995) Photosynthetic responses to temperature, light flux-density, CO₂ concentration and vapour pressure deficit in *Eucalyptus tetrodonta* grown under CO₂ enrichment. *Environmental Pollution* **90**, 41–49.
- Eamus D. & Jarvis P.G. (1989) The direct effects of increase in the global atmospheric CO₂ concentration on natural and commercial temperate trees and forests. *Advances in Ecological Research* **19**, 1–55.
- Egli P. & Körner Ch. (1997) Growth responses to elevated CO₂ and soil quality in beech–spruce model ecosystems. *Acta Oecologia* **18**, 343–349.
- El Kohen A., Venet L. & Mousseau M. (1993) Growth and photosynthesis of two deciduous forest tree species exposed to elevated carbon dioxide. *Functional Ecology* **7**, 480–486.
- Ellsworth D.S., Oren R., Huang C., Phillips N. & Hendrey G.R. (1995) Leaf and canopy responses to elevated CO₂ in a pine forest under free-air CO₂ enrichment. *Oecologia* **104**, 139–146.
- Epron D., Liozon R. & Mousseau M. (1996) Effects of elevated CO₂ concentration on leaf characteristics and photosynthetic capacity of beech (*Fagus sylvatica*) during the growing season. *Tree Physiology* **16**, 425–432.
- Field C.B., Jackson R.B. & Mooney H.A. (1995) Stomatal responses to increased CO₂: implications from the plant to global scale. *Plant, Cell and Environment* **18**, 1214–1225.
- Field C.B. & Mooney H.A. (1986) The photosynthesis–nitrogen relationship in wild plants. In *On the Economy of Plant Form and Function* (ed. T.J. Givnish), pp. 25–55. Cambridge University Press, Cambridge.
- Gahrooei F.R. (1998) Impacts of elevated atmospheric CO₂ on

- litter quality, litter decomposability and nitrogen turnover rate of two oak species in a Mediterranean forest ecosystem. *Global Change Biology* **4**, 667–677.
- Garcia R.L., Idso S.B., Wall G.W. & Kimball B.A. (1994) Changes in net photosynthesis and growth of *Pinus elliottii* seedlings in response to atmospheric CO₂ enrichment. *Plant, Cell and Environment* **17**, 971–978.
- Goodfellow J., Eamus D. & Duff G. (1997) Diurnal and seasonal changes in the impact of CO₂ enrichment on assimilation, stomatal conductance and growth in a long-term study of *Mangifera indica* in the wet-dry tropics of Australia. *Tree Physiology* **17**, 291–299.
- Goulden M.L., Munger J.W., Fan S.M., Daube B.C. & Wofsy S.C. (1996) Exchange of carbon dioxide by a deciduous forest: response to interannual climate variability. *Science* **271**, 1576–1578.
- Gregory K.M. (1996) Are paleoclimate estimates biased by foliar physiognomic responses to increased atmospheric CO₂? *Palaeogeography Palaeoclimatology Palaeoecology* **124**, 39–51.
- Griffin K.L., Ball J.T. & Strain B.R. (1996a) Direct and indirect effects of elevated CO₂ on whole-shoot respiration in ponderosa pine seedlings. *Tree Physiology* **16**, 33–41.
- Griffin K.L., Winner W.E. & Strain B.R. (1996b) Construction cost of loblolly and ponderosa pine leaves grown with varying carbon and nitrogen availability. *Plant, Cell and Environment* **19**, 729–738.
- Guak S., Olszyk D.M., Fuchigami L.H. & Tingey D.T. (1998) Effects of elevated CO₂ and temperature on cold hardiness and spring bud burst and growth in Douglas-fir (*Pseudotsuga menziesii*). *Tree Physiology* **18**, 671–679.
- Gunderson C.A., Norby R.J. & Wullschlegel S.D. (1993) Foliar gas exchange responses of two deciduous hardwoods during 3 years of growth in elevated CO₂: no loss of photosynthetic enhancement. *Plant, Cell and Environment* **16**, 797–807.
- Gunderson C.A. & Wullschlegel S.D. (1994) Photosynthetic acclimation of forest trees to a doubling of atmospheric CO₂: a broader perspective. *Photosynthesis Research* **39**, 369–388.
- Hättenschwiler S. & Körner Ch. (1998) Biomass allocation and canopy development in spruce model ecosystems under elevated CO₂ and increased N deposition. *Oecologia* **113**, 104–114.
- Hättenschwiler S., Miglietta F., Raschi A. & Körner Ch. (1997a) Thirty years of *in situ* tree growth under elevated CO₂: a model for future forest responses? *Global Change Biology* **3**, 463–471.
- Hättenschwiler S., Miglietta F., Raschi A. & Körner Ch. (1997b) Morphological adjustments of mature *Quercus ilex* trees to elevated CO₂. *Acta Oecologia* **18**, 361–365.
- Henderson-Sellers A., McGuffie K. & Gross C. (1995) Sensitivity of global climate model simulations to increased stomata resistance and CO₂ increases. *Journal of Climate* **8**, 1738–1756.
- Hogan K.P., Fleck I., Bungard R., Cheeseman J.M. & Whitehead D. (1997) Effects of elevated CO₂ on the utilization of light energy in *Nothofagus fusca* and *Pinus radiata*. *Journal of Experimental Botany* **48**, 1289–1297.
- Hogan K.P., Whitehead D., Kallarackal J., Buwalda J.G., Meekings J. & Rogers G.N.D. (1996) Photosynthetic activity of leaves of *Pinus radiata* and *Nothofagus fusca* after 1 year of growth at elevated CO₂. *Australian Journal of Plant Physiology* **23**, 623–630.
- Idso S.B. (1991) The aerial fertilization effect of CO₂ and its implications for global carbon cycling and maximum greenhouse warming. *Bulletin American Meteorological Society* **72**, 962–965.
- Idso K.E. & Idso S.B. (1994) Plant responses to atmospheric CO₂ enrichment in the face of environmental constraints: a review of the past 10 years research. *Agricultural and Forest Meteorology* **69**, 153–203.
- Idso S.B., Idso K.E., Garcia R.L., Kimball B.A. & Hooper J.K. (1995) Effects of atmospheric CO₂ enrichment and foliar methanol application of net photosynthesis of sour orange (*Citrus aurantium*; Rutaceae) leaves. *American Journal of Botany* **82**, 26–30.
- Idso S.B. & Kimball B.A. (1991) Downward regulation of photosynthesis and growth at high CO₂ levels. No evidence for either phenomenon in three-year study of sour orange trees. *Plant Physiology* **96**, 990–992.
- Idso S.B. & Kimball B.A. (1992a) Effects of atmospheric CO₂ enrichment on photosynthesis, respiration, and growth of sour orange trees. *Plant Physiology* **99**, 341–343.
- Idso S.B. & Kimball B.A. (1992b) Seasonal fine-root biomass development of sour orange trees grown in atmospheres of ambient and elevated CO₂ concentration. *Plant, Cell and Environment* **15**, 337–341.
- Idso S.B. & Kimball B.A. (1993) Tree growth in carbon dioxide enriched air and its implications for global carbon cycling and maximum levels of atmospheric CO₂. *Global Biogeochemical Cycles* **7**, 537–555.
- Idso S.B. & Kimball B.A. (1994) Effects of atmospheric CO₂ enrichment on biomass accumulation and distribution in Eldarica pine trees. *Journal of Experimental Botany* **45**, 1669–1672.
- Idso S.B. & Kimball B.A. (1997) Effects of long-term atmospheric CO₂ enrichment on the growth and fruit production of sour orange trees. *Global Change Biology* **3**, 89–96.
- Idso S.B., Kimball B.A., Akin D.E. & Kridler J. (1993b) A general relationship between carbon dioxide-induced reductions in stomatal conductance and concomitant increases in foliage temperature. *Environmental and Experimental Botany* **33**, 443–446.
- Idso S.B., Kimball B.A. & Allen S.G. (1991) Net photosynthesis of sour orange trees maintained in atmospheres of ambient and elevated CO₂ concentration. *Agricultural and Forest Meteorology* **54**, 95–101.
- Idso S.B., Kimball B.A. & Hendrix D.L. (1993a) Air-temperature modifies the size-enhancing effects of atmospheric CO₂ enrichment on sour orange tree leaves. *Environmental and Experimental Botany* **33**, 293–299.
- Idso S.B., Wall G.W. & Kimball B.A. (1993c) Interactive effects of atmospheric CO₂ enrichment and light intensity reductions on net photosynthesis of sour orange tree leaves. *Environmental and Experimental Botany* **33**, 367–373.
- Ineson P. & Cotrufo M.F. (1997) Increasing concentrations of atmospheric CO₂ and decomposition processes in forest ecosystems. In *Plant Responses to Elevated CO₂. Evidence from Natural Springs* (eds A. Raschi, F. Miglietta, R. Tognetti & P.R. van Gardingen.), pp. 242–267. Cambridge University Press, Cambridge.
- Janssens I.A., Crookshanks M., Taylor G. & Ceulemans R. (1998) Elevated atmospheric CO₂ increases fine root production, respiration, rhizosphere respiration and soil CO₂ efflux in young Scots pine seedlings. *Global Change Biology* **4**, 871–878.
- Jarvis P.G. (1995) Scaling processes and problems. *Plant, Cell and Environment* **18**, 1079–1089.
- Jarvis P.G. (1998) Effects of climate change on ecosystem carbon balance. In *The Earth's Changing Land, GCTE-LUCC Open Science Conference on Global Change, Abstracts* p. 198. Institut Cartografic de Catalunya, Barcelona.
- Johnson D.W., Ball J.T. & Walker R.F. (1997) Effects of CO₂ and nitrogen fertilization on vegetation and soil nutrient content in juvenile ponderosa pine. *Plant and Soil* **190**, 29–40.
- Johnson D., Henderson P.H., Ball J.T. & Walker R.F. (1996) Effects of CO₂ and N on growth and N dynamics in ponderosa pine: results from the first two growing seasons. In *Carbon Dioxide and Terrestrial Ecosystems* (eds G.W. Koch & H.A. Mooney), pp. 23–40. Academic Press, San Diego.
- Karnosky D.F., Podila G.K., Gagnon Z., Pechter P., Akkapeddi A., Sheng Y., Riemenschneider D.E., Coleman M.D., Dickson R.E.

- & Isebrands J.G. (1998) Genetic control of responses to interacting tropospheric ozone and CO₂ in *Populus tremuloides*. *Chemosphere* **36**, 807–812.
- Kellomäki S. & Wang K.-Y. (1996) Photosynthetic responses to needle water potentials in Scots pine after a four-year exposure to elevated CO₂ and temperature. *Tree Physiology* **16**, 765–772.
- Kellomäki S. & Wang K.-Y. (1997a) Effects of long-term CO₂ and temperature elevation on crown nitrogen distribution and daily photosynthetic performance of Scots pine. *Forest Ecology and Management* **99**, 309–326.
- Kellomäki S. & Wang K.-Y. (1997b) Effects of elevated O₃ and CO₂ concentrations on photosynthesis and stomatal conductance in Scots pine. *Plant, Cell and Environment* **20**, 995–1006.
- Körner Ch. (1995) Towards a better experimental basis for upscaling plant responses to elevated CO₂ and climate warming. *Plant, Cell and Environment* **18**, 1101–1110.
- Körner Ch. & Arnone J.A. (1992). Responses to elevated carbon dioxide in artificial tropical ecosystems. *Science* **257**, 1672–1675.
- Körner Ch. & Miglietta F. (1994) Long term effects of naturally elevated CO₂ on mediterranean grassland and forest trees. *Oecologia* **99**, 343–351.
- Kramer P.J. (1981) Carbon dioxide concentration, photosynthesis, and dry matter production. *Bioscience* **31**, 29–33.
- Kubiske M.E. & Pregitzer K.S. (1996) Effects of elevated CO₂ and light availability on the photosynthetic light response of trees of contrasting shade tolerance. *Tree Physiology* **16**, 351–358.
- Kubiske M.E., Pregitzer K.S., Mikan C.J., Zak D.R., Maziasz J.L. & Teeri J.A. (1997) *Populus tremuloides* photosynthesis and crown architecture in response to elevated CO₂ and soil N availability. *Oecologia* **110**, 328–336.
- Le Thiec D., Dixon M., Loosveldt P. & Garrec J.P. (1995) Seasonal and annual variations of phosphorus, calcium, potassium and manganese contents in different cross-sections of *Picea abies* (L.) Karst. needles and *Quercus rubra* L. leaves exposed to elevated CO₂. *Trees* **10**, 55–62.
- Lee H.S.J. & Jarvis P.G. (1995) Trees differ from crops and each other in their responses to increases in CO₂ concentration. *Journal of Biogeography* **22**, 323–330.
- Lee H., Overdieck D. & Jarvis P.G. (1998) Biomass, growth and carbon allocation. In *European Forests and Global Change: Likely Impacts of Rising CO₂ and Temperature* (ed. P.G. Jarvis), chapter 5. Cambridge University Press, Cambridge, UK.
- Lewis J.D., Tissue D.T. & Strain B.R. (1996) Seasonal response of photosynthesis to elevated CO₂ in loblolly pine (*Pinus taeda* L.) over 2 growing seasons. *Global Change Biology* **2**, 103–114.
- Lincoln D.E., Fajer E.D. & Jonson R.H. (1993) Plant–insect herbivore interactions in elevated CO₂ environments. *Trends in Ecology and Evolution* **8**, 64–68.
- Lindroth R.L., Roth S., Kruger E.L., Volin J.C. & Koss P.A. (1997) CO₂-mediated changes in aspen chemistry: effects on gypsy moth performance and susceptibility to virus. *Global Change Biology* **3**, 279–289.
- Long S.P. (1991) Modification of the response of photosynthetic productivity to rising temperature by atmospheric CO₂ concentrations: Has its importance been underestimated? *Plant, Cell and Environment* **14**, 729–739.
- Long S.P. & Drake B.G. (1991) Effect of the long term elevation of CO₂ concentration in the field on the quantum yield of photosynthesis of the C₃ sedge, *Scirpus olneyi*. *Plant Physiology* **96**, 221–226.
- Marek M.V., Kalina J. & Matoušková M. (1995) Response of photosynthetic carbon assimilation of Norway spruce exposed to long-term elevation of CO₂ concentration. *Photosynthetica* **31**, 209–220.
- Martin P. (1992) EXE: a climatically sensitive model to study climate change and CO₂ enhancement effects on forest. *Australian Journal of Botany* **40**, 717–735.
- McGuire A.D., Melillo J.M. & Joyce L.A. (1995) The role of nitrogen in the response of forest net primary production to elevated atmospheric carbon dioxide. *Annual Review of Ecology and Systematics* **26**, 473–503.
- McGuire A.D., Melillo J.M., Kicklighter D.W., Pan Y., Xiao X., Helfrich J., Moore I.I.B., Vorosmarty J. & Schloss A.L. (1997) Equilibrium responses of global net primary production and carbon storage to doubled atmospheric carbon dioxide: Sensitivity to changes in vegetation nitrogen concentration. *Global Biogeochemical Cycles* **11**, 173–189.
- Miglietta F., Raschi A., Bettarini I., Resti R. & Selvi F. (1993) Natural CO₂ springs in Italy: a resource for examining long-term response of vegetation to rising atmospheric CO₂ concentrations. *Plant, Cell and Environment* **16**, 873–878.
- Mooney H.A., Canadell J., Chapin F.S., Ehleringer J., Körner Ch., McMurtrie R., Parton W.J., Pitelka L. & Schulze E.-D. (1999) Ecosystem physiology responses to global change. In *The Terrestrial Biosphere and Global Change. Implications for Natural and Managed Ecosystems* (eds B.H. Walker, W.L. Steffen, J. Canadell and J.S.I. Ingram), pp. 141–189. Cambridge University Press, Cambridge.
- Morison J.I.L. (1985) Sensitivity of stomata and water use efficiency to high CO₂. *Plant, Cell and Environment* **8**, 467–474.
- Mousseau M., Dufrene E., El Kohen A., Epron D., Godard D., Liozon R., Pontailler J.Y. & Saugier B. (1996) Growth strategy and tree response to elevated CO₂: a comparison of beech (*Fagus sylvatica*) and sweet chestnut (*Castanea sativa* Mill.) In *Carbon Dioxide and Terrestrial Ecosystems* (eds G.W. Koch & H.A. Mooney), pp. 71–86. Academic Press, San Diego.
- Murray M.B. & Ceulemans R. (1998). Will tree foliage be larger and live longer? In *European Forests and Global Change: Likely Impacts of Rising CO₂ and Temperature* (ed. P.G. Jarvis), pp. 94–125. Cambridge University Press, Cambridge.
- Murray M.B., Smith R.I., Leith I.D., Fowler D., Lee H.S.J., Friend A.D. & Jarvis P.G. (1994) Effects of elevated CO₂, nutrition and climatic warming on bud phenology in Sitka spruce (*Picea sitchensis*) and their impact on the risk of frost damage. *Tree Physiology* **14**, 691–706.
- Norby R.J. (1994) Issues and perspectives for investigating root responses to elevated atmospheric carbon dioxide. *Plant and Soil* **165**, 9–20.
- Norby R.J. (1996) Forest canopy productivity index. *Nature* **381**, 564.
- Norby R.J. (1998) Nitrogen deposition: a component of global change analyses. *New Phytologist* **139**, 189–200.
- Norby R.J. & O'Neill E.G. (1989) Growth dynamics and water use of seedlings of *Quercus alba* L. in CO₂-enriched atmospheres. *New Phytologist* **111**, 491–500.
- Norby R.J. & O'Neill E.G. (1991) Leaf area compensation and nutrient interactions in CO₂-enriched yellow-poplar (*Liriodendron tulipifera* L.) seedlings. *New Phytologist* **117**, 515–528.
- Norby R.J. & Sigal L.L. (1989) Nitrogen fixation in the lichen *Lobaria pulmonaria* in elevated atmospheric carbon dioxide. *Oecologia* **79**, 566–568.
- Norby R.J., O'Neill E.G. & Luxmoore R.G. (1986) Effects of atmospheric CO₂ enrichment on the growth and mineral nutrition of *Quercus alba* seedlings in nutrient-poor soil. *Plant Physiology* **82**, 83–89.
- Norby R.J., Edwards N.T., Riggs J.S., Abner C.H., Wullschlegel S.D. & Gunderson C.A. (1997) Temperature-controlled open-top chambers for global change research. *Global Change Biology* **3**, 259–267.
- Norby R.J., Gunderson C.A., Wullschlegel S.D., O'Neill E.G. & McCracken M.K. (1992) Productivity and compensatory response of yellow poplar trees in elevated CO₂. *Nature* **357**, 322–324.
- Norby R.J., Verbrugge M.J., Hartz J.S., Wullschlegel S.D., Gunderson C.A., O'Neill E.G. & Edwards N.T. (1998) Increased temperature has both positive and negative influences on tree

- growth. In *The Earth's Changing Land, GCTE-LUCC Open Science Conference on Global Change, Abstracts*, p. 24. Institut Cartografic de Catalunya, Barcelona.
- Norby R.J., Wullschlegel S.D. & Gunderson C.A. (1996) Tree responses to elevated CO₂ and the implications for forests. In *Carbon Dioxide and Terrestrial Ecosystems* (eds G.W. Koch & H.A. Mooney), pp. 1–21. Academic Press, San Diego.
- Norby R.J., Wullschlegel S.D., Gunderson C.A. & Nietch C.T. (1995) Increased growth efficiency of *Quercus alba* trees in a CO₂-enriched atmosphere. *New Phytologist* **131**, 91–97.
- O'Neill E.G. & Norby R.J. (1996) Litter quality and decomposition rates of foliar litter produced under CO₂ enrichment. In *Carbon Dioxide and Terrestrial Ecosystems* (eds G.W. Koch & H.A. Mooney), pp. 87–103. Academic Press, San Diego.
- Oechel W.C. & Strain B.R. (1985) Native species responses to increased atmospheric carbon dioxide concentration. In *Direct Effects of Increasing Carbon Dioxide on Vegetation* (eds B.R. Strain & J.D. Cure), pp. 117–154. DOE/ER-0238. U.S. Department of Energy, Washington, D.C.
- Overdieck D. (1993) Effects of atmospheric CO₂ enrichment on CO₂ exchange rates of beech stands in small model ecosystems. *Water, Soil and Air Pollution* **70**, 259–277.
- Paul E.A. & Clark F.E. (1989) *Soil Microbiology and Biochemistry*. Academic Press, New York.
- Peñuelas J., Idso S.B., Ribas A. & Kimball B.A. (1997) Effects of long-term atmospheric CO₂ enrichment on the mineral concentration of *Citrus aurantium* leaves. *New Phytologist* **135**, 439–444.
- Pettersson R. & McDonald A.J.S. (1994) Effects of nitrogen supply on acclimation of photosynthesis to elevated CO₂. *Photosynthesis Research* **39**, 389–400.
- Pollard D. & Thompson S.L. (1995) Use of a land-surface scheme (LSX) in a global climate model — the response to doubling stomatal resistance. *Global and Planetary Change* **10**, 129–161.
- Poorter H., Roumet C. & Campbell B.D. (1996) Interspecific variation in the growth response of plants to elevated CO₂: a search for functional types. In *Carbon Dioxide, Populations, and Communities* (eds Ch. Körner & F.A. Bazzaz), pp. 375–412. Academic Press, San Diego.
- Pregitzer K.S., Zak D.R., Curtis P.S., Kubiske M.E., Teeri J.A. & Vogel C.S. (1995) Atmospheric CO₂, soil nitrogen and turnover of fine roots. *New Phytologist* **129**, 579–585.
- Pritchard S., Peterson C., Runion G.B., Prior S. & Rogers H. (1997) Atmospheric CO₂ concentration, N availability, and water status affect patterns of ergastic substance deposition in longleaf pine (*Pinus palustris* Mill.) foliage. *Trees* **11**, 494–503.
- Randlett D.L., Zak D.R., Pregitzer K.S. & Curtis P.S. (1996) Elevated atmospheric carbon dioxide and leaf litter chemistry: influences on microbial respiration and net nitrogen mineralization. *Soil Science Society of America Journal* **60**, 1571–1577.
- Rastetter E.B., McKane R.B., Shaver G.R. & Melillo J.M. (1992) Changes in C storage by terrestrial ecosystems: how C–N interactions restrict responses to CO₂ and temperature. *Water, Air and Soil Pollution* **64**, 327–344.
- Rey A. & Jarvis P.G. (1997) Growth response of young birch trees (*Betula pendula* Roth.) after four and a half years of CO₂ exposure. *Annals of Botany* **80**, 809–816.
- Rey A. & Jarvis P.G. (1998) Long-term photosynthetic acclimation to increased atmospheric CO₂ concentration in young birch (*Betula pendula*) trees. *Tree Physiology* **18**, 441–450.
- Rogers H.H., Heck W.W. & Heagle A.S. (1983) A field technique for the study of plant responses to elevated carbon dioxide concentrations. *Journal of the Air Pollution Control Association* **33**, 42–44.
- Roth S., Lindroth R.L., Volin J.C. & Kruger E.L. (1998) Enriched atmospheric CO₂ and defoliation: effects on tree chemistry and insect performance. *Global Change Biology* **4**, 419–430.
- Ruimy A., Field C.B., Herbert D., Kelly R., McMurtrie R.E., Parton W.J., Pierce L.L. & CMEAL participants (1999) Forest and grassland responses to elevated atmospheric CO₂: resource use factors from four ecosystem models. *Ecological Applications*, in press.
- Runion G.B., Mitchell R.J., Rogers H.H., Prior S.A. & Counts T.K. (1997) Effects of nitrogen and water limitation and elevated atmospheric CO₂ on ectomycorrhiza of longleaf pine. *New Phytologist* **137**, 681–689.
- Rygiewicz P.T., Johnson M.G., Ganio L.M., Tingey D.T. & Storm M.J. (1997) Lifetime and temporal occurrence of ectomycorrhizae on ponderosa pine (*Pinus ponderosa* Laws.) seedlings grown under varied atmospheric CO₂ and nitrogen levels. *Plant and Soil* **189**, 275–287.
- Sage R.F. (1994) Acclimation of photosynthesis to increasing atmospheric CO₂: The gas exchange perspective. *Photosynthesis Research* **39**, 351–368.
- Saxe H., Ellsworth D.S. & Heath J. (1998) Tansley review no. 98. Tree and forest functioning in an enriched CO₂ atmosphere. *New Phytologist* **139**, 395–436.
- Scarascia-Mugnozza G., De Angelis P., Matteucci G. & Valentini R. (1996) Long-term exposure to elevated [CO₂] in a natural *Quercus ilex* L. community: net photosynthesis and photochemical efficiency of PSII at different levels of water stress. *Plant, Cell and Environment* **19**, 643–654.
- Sellers P.J., Bounoua L., Collatz G.J., Randall D.A., Dazlich D.A., Los S.O., Berry J.A., Fung I., Tucker C.J., Field C.B. & Jensen T.G. (1996) Comparison of radiative and physiological effects of doubled atmospheric CO₂ on climate. *Science* **271**, 1402–1406.
- Strain B.R. (1985) Physiological and ecological controls on carbon sequestering in terrestrial ecosystems. *Biogeochemistry* **1**, 219–232.
- Stulen I. & den Hertog J. (1993) Root growth and functioning under atmospheric CO₂ enrichment. *Vegetatio* **104/105**, 99–115.
- Surano K.A., Daley P.F., Houppis J.L.J., Shinn J.H., Helms J.A., Palassou R.J. & Costella M.P. (1986) Growth and physiological responses of *Pinus ponderosa* Dougl. ex P. Laws. to long-term elevated CO₂ concentration. *Tree Physiology* **2**, 243–259.
- Teskey R.O. (1995) A field study of the effects of elevated CO₂ on carbon assimilation, stomatal conductance and leaf and branch growth of *Pinus taeda* trees. *Plant, Cell and Environment* **18**, 565–573.
- Thomas S.M., Whitehead D., Adams J.A., Reid J.B., Sherlock R.R. & Leckie A.C. (1996) Seasonal root distribution and soil surface carbon fluxes for one-year-old *Pinus radiata* trees growing at ambient and elevated carbon dioxide concentration. *Tree Physiology* **16**, 1015–1021.
- Tingey D.T., Johnson M.G., Phillips D.L., Johnson D.W. & Ball J.T. (1996) Effects of elevated CO₂ and nitrogen on the synchrony of shoot and root growth in ponderosa pine. *Tree Physiology* **16**, 905–914.
- Tingey D.T., Phillips D.L., Johnson M.G., Storm M.J. & Ball J.T. (1997) Effects of elevated CO₂ and nitrogen on fine root dynamics and fungal growth in seedling *Pinus ponderosa*. *Environmental and Experimental Botany* **37**, 73–83.
- Tissue D.T., Griffin K.L. & Ball J.T. Photosynthetic adjustment in field-grown ponderosa pine trees after six years exposure to elevated CO₂. *Tree Physiology*, in press.
- Tissue D.T., Thomas R.B. & Strain B.R. (1993) Long-term effects of elevated CO₂ and nutrients on photosynthesis and rubisco in loblolly-pine seedlings. *Plant, Cell and Environment* **16**, 859–865.
- Tissue D.T., Thomas R.B. & Strain B.R. (1996) Growth and photosynthesis of loblolly pine (*Pinus taeda*) after exposure to elevated CO₂ for 19 months in the field. *Tree Physiology* **16**, 49–59.
- Tissue D.T., Thomas R.B. & Strain B.R. (1997) Atmospheric CO₂ enrichment increases growth and photosynthesis in *Pinus taeda*: a 4 year experiment in the field. *Plant, Cell and Environment* **20**, 1123–1134.

- Tognetti R., Giovannelli A., Longobucco A., Miglietta F. & Raschi A. (1996) Water relations of oak species growing in the natural CO₂ spring of Rapolano (central Italy). *Annales Des Sciences Forestières* **53**, 475–485.
- Turnbull M.H., Tissue D.T., Griffin K.L., Rogers G.N.D. & Whitehead D. (1998) Photosynthetic acclimation to long-term exposure to elevated CO₂ concentration in *Pinus radiata* Don. is related to age of needles. *Plant, Cell and Environment* **21**, 1019–1028.
- Turner J. (1977) Effect of nitrogen availability on nitrogen cycling in a Douglas-fir stand. *Forest Science* **23**, 307–316.
- Vogel C.S. & Curtis P.S. (1995) Leaf gas exchange and nitrogen dynamics of N₂-fixing field-grown *Alnus glutinosa* under elevated atmospheric CO₂. *Global Change Biology* **1**, 55–61.
- Walker R.F., Geisinger D.R., Johnson D.W. & Ball J.T. (1995) Interactive effects of atmospheric CO₂ enrichment and soil N on growth and ectomycorrhizal colonization of ponderosa pine seedlings. *Forest Science* **41**, 491–500.
- Walker R.F., Geisinger D.R., Johnson D.W. & Ball J.T. (1997) Elevated atmospheric CO₂ and soil N fertility effects on growth, mycorrhizal colonization, and xylem water potential of juvenile ponderosa pine in a field soil. *Plant and Soil* **195**, 25–36.
- Wang K.-Y. & Kellomäki S. (1997) Stomatal conductance and transpiration in shoots of Scots pine after 4-year exposure to elevated CO₂ and temperature. *Canadian Journal of Botany* **75**, 552–561.
- Wang K.-Y., Kellomäki S. & Laitinen K. (1995) Effects of needle age, long-term temperature and CO₂ treatments on the photosynthesis of Scots pine. *Tree Physiology* **15**, 211–218.
- Wang Y.-P., Rey A. & Jarvis P.G. (1998) Carbon balance of young birch trees grown in ambient and elevated atmospheric CO₂ concentrations. *Global Change Biology* **4**, 797–807.
- Waring R.H. & Schlesinger W.H. (1985) *Forest Ecosystems. Concepts and Management*. Academic Press, Orlando.
- Will R.E. & Ceulemans R. (1997) Effects of elevated CO₂ concentration on photosynthesis, respiration and carbohydrate status of coppice *Populus* hybrids. *Physiologia Plantarum* **100**, 933–939.
- Williams R.S., Lincoln D.E. & Norby R.J. (1998) Leaf age effects of elevated CO₂-grown white oak leaves on spring-feeding lepidopterans. *Global Change Biology* **4**, 235–246.
- Williams R.S., Lincoln D.E. & Thomas R.B. (1997) Effects of elevated CO₂-grown loblolly pine needles on the growth, consumption, and pupal weight of red-headed pine sawfly reared within open-topped chambers. *Global Change Biology* **3**, 501–511.
- Wong S.-C. & Dunin F.X. (1987) Photosynthesis and transpiration of trees in a eucalypt forest stand: CO₂, light and humidity responses. *Australian Journal of Plant Physiology* **14**, 619–632.
- Woodward F.I., Smith T.M. & Emanuel W.R. (1995) A global land primary productivity and phytogeography model. *Global Biogeochemical Cycles* **9**, 471–490.
- Wullschleger S.D. & Norby R.J. (1992) Respiratory cost of leaf growth and maintenance in white oak saplings exposed to atmospheric CO₂ enrichment. *Canadian Journal of Forest Research* **22**, 1717–1721.
- Wullschleger S.D., Norby R.J. & Gunderson C.A. (1992a) Growth and maintenance respiration in leaves of *Liriodendron tulipifera* L. exposed to long-term carbon dioxide enrichment in the field. *New Phytologist* **121**, 515–523.
- Wullschleger S.D., Norby R.J. & Gunderson C.A. (1997a) Forest trees and their response to atmospheric CO₂ enrichment: A compilation of results. In *Advances in Carbon Dioxide Effects Research* (eds L.H. Allen Jr, M.B. Kirkham, D.M. Olszyk & C.E. Williams), pp. 79–100. ASA Special Publication no. 61, American Society of Agronomy, Madison, WI.
- Wullschleger S.D., Norby R.J. & Hanson P.J. (1995b) Growth and maintenance respiration in stems of *Quercus alba* after four years of CO₂ enrichment. *Physiologia Plantarum* **93**, 47–54.
- Wullschleger S.D., Norby R.J. & Hendrix D.L. (1992b) Carbon exchange rates, chlorophyll content, and carbohydrate status of two forest tree species exposed to carbon dioxide enrichment. *Tree Physiology* **10**, 21–31.
- Wullschleger S.D., Norby R.J., Love J.C. & Runck C. (1997b) Energetic costs of tissue construction in yellow-poplar and white oak trees exposed to long-term CO₂ enrichment. *Annals of Botany* **80**, 289–297.
- Wullschleger S.D., Post W.M. & King A.W. (1995a). On the potential of a CO₂ fertilization effect in forests: Estimates of the biotic growth factor based on 58 controlled-exposure studies. In *Biotic Feedbacks in the Global Climatic System: Will Warming Feed the Warming?* (eds G.M. Woodwell & F.T. Mackenzie), pp. 85–107. Oxford University Press, New York.
- Zak D.R., Pregitzer K.S., Curtis P.S., Teeri J.A., Fogel R. & Randlett D.L. (1993) Elevated atmospheric CO₂ and feedback between carbon and nitrogen cycles. *Plant and Soil* **151**, 105–117.